



## Prospects of Probiotics and Fish Growth-promoting Bacteria in Aquaculture: A Review

Rekha Kalita<sup>1,2,3</sup>, Amrit Pegu<sup>1,2</sup> and Chittaranjan Baruah<sup>1\*</sup>

<sup>1</sup>Postgraduate Department of Zoology, Darrang College (Gauhati University), Tezpur- 784001, Assam, India

<sup>2</sup>CSIR-Junior Research Fellow, Department of Zoology, Gauhati University, Guwahati-781014, Assam, India

<sup>3</sup>Department of Zoology, Behali Degree College (Gauhati University), Borgang, Biswanath-784167, Assam, India

\*Corresponding author: [chittaranjan.baruah@gmail.com](mailto:chittaranjan.baruah@gmail.com)

### ABSTRACT

Modern aquaculture faces significant obstacles including a deficiency in sources of protein in the feed, vulnerability to infections, and quality degradation during growing and storage, despite being the sector with the highest growth rate. Beneficial bacterial species shield aquatic animals from infection or stop product deterioration, and bacterial biomass is thought to be a potential source of protein for animal feed. This review focuses on the nutritional, anti-pathogenic, and immunoregulatory functions of these bacteria in relation to aquatic products. Additionally, we examine the connection between host immunity, beneficial bacteria, and gut microbiota, along with recent advances in our understanding of host immunity and microbial mutualism. This analysis emphasizes specific microbial metabolites, bacterial components, and the immune system. The actions of probiotics in aquatic animals have garnered significant research attention in recent years. Various findings, with the potential to be both academically innovative and practically helpful, have emerged. The positive effects of beneficial bacteria on aquatic organisms have sparked extensive research, wide-ranging use in aquaculture, and innovative applications. Future improvements to current practices will require the development of novel applications and related mechanistic research.

**Keywords:** Probiotics, Growth-promotion, Fish, Vaccine, Immunoregulation.

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### INTRODUCTION

The aquaculture industry was developed and is currently the fastest-growing to meet the increasing demand for fish protein and represents one-third of world fisheries production (FAO, 2020). With the increasing population, the demand for aquatic food has also increased, and this can be fulfilled by the multibillion-dollar, rapidly expanding aquaculture industry. Of the total production, 63 percent (112 million metric tons) was harvested in marine waters (70 percent from capture fisheries and 30 percent from aquaculture) and 37 percent (66 million metric tons) in inland waters (83 percent from aquaculture and 17 percent from capture fisheries) (FAO, 2022). All over the world, per year, the fish production industry has been gradually growing at a standard compounded rate of 9.2% since 1970 (El-Saadony et al., 2021). The aquaculture industry has partly compensated for the reduction of capture fisheries, and many reports show that it will be the future weapon for getting rid of this problem (El-Saadony et al., 2021). By 2050, aquaculture production is anticipated to double, allowing it to meet demand while relieving the strain on wild fishing (Yukgehnaish et al., 2020). According to projections by the United Nations Food and Agriculture Organization, farm-raised fish will contribute two-thirds of the world's consumption of seafood by the year 2030 (Thorpe and Castillo, 2018).

The major constraint is the presence of pathogenic microorganisms. Disease caused by this harmful microbiota creates challenges for farmers and finally impacts net production. Although various kinds of vaccines are being developed by experts, they are not the universal solution. Therefore, the management of disease in aquaculture became a very crucial problem for farmers. Later, to solve this problem, antibiotics are used as a traditional strategy for a long period of time. According to many scientific reports, antibiotics are able to control many disease-causing bacteria and can improve fish health in aquaculture practices (El-Saadony et al., 2021). But, as we know, each coin has two sides. Likewise, everything has two effects. Antibiotics kill some beneficial bacteria, which are inhabitants of the fish gastrointestinal tract. However, it also caused fish products to acquire antibiotic residues that are unsafe for human consumption (WHO, 2006). These concerns, along with the detrimental impact of products and residual antibiotics on human health, led the European Union and the United States to impose prohibitions on the use of antibiotics (Kesarcodi-Watson et al., 2008).

With the ban on antibiotics, there was a need for a new management strategy for fish health. To fulfill this need, fishery scientists have conducted much research and they came to an unbeatable conclusion on the use of probiotics. Later, this new idea has received much more attention and has been recognized as a natural disease controller and a good growth-promoting agent for fish in aquaculture

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practice. The Greek words "pro" and "bios" gave rise to the phrase "probiotic," which means "for life" (Dailey et al., 2019). The term "probiotic" refers to a material that can promote the growth of other organisms (Thatcher et al., 2022).

Antimicrobial drugs and growth promoters are being replaced by these beneficial microorganisms. Fish pathogenic bacteria and probiotic bacteria compete for nutrients and sites of attachment in the stomach, adhering there to prevent pathogen growth, as some candidate probiotics have been shown to produce antibacterial substances that inhibit the growth of harmful microbes and maintain intestinal microecological balance (Xu et al., 2014). Thus, several probiotics used in aquaculture have been documented to exert direct antibacterial activities against known pathogens. Several experiments concluded that beneficial bacteria could enhance aquaculture production along with the disease-causing bacteria in aquaculture (Allameh et al., 2017).

Lactic acid bacteria (LAB) are the major group of microorganisms used as probiotics for animals. In the fish production industry, LABs enhance resistance to pathogens, survivability, feed conversion efficiency, protein efficiency ratio, and digestibility. They also prevent intestinal disorders and can neutralize the anti-nutritional factors present in feedstuffs. In addition, LABs improve the fish immune system. LABs are those that can improve bacterial growth and monitoring when used in aquaculture (Allameh et al., 2017). Probiotic-fortified meal increases the metabolic process of the fish body and helps it grow in size.

#### Mechanism of Action of Bacteria for Fish Growth Promotion

Microorganisms inhabiting fish internal tissues play a variety of crucial roles, including digestion, immunity, defense against harmful bacteria, and other developments. Govindaraj et al. (2021) isolated 120 beneficial probiotics from the intestines of freshwater fishes: *Cirrhinus mrigala*, *Puntius filamentosus*, *Channa striata*, *Rasbora daniconius* and *Oreochromis mossambicus*. Among these, seven probiotics have been found to have strong antagonistic activity against several fish pathogens. They concluded that *Lactococcus fermentum* has high probiotic potential and is an effective dietary supplement for modern freshwater aquaculture. In their study, *L. fermentum* isolated from *C. mrigala* showed different extracellular enzyme secretions activity like amylase, lipase, and protease. Moreover, *in vitro* evaluations of intestinal mucus indicate that *L. fermentum* displayed potential adherence capacity and an increased survival rate after being inoculated with *Artemia nauplii*.

Banerjee et al. (2013) investigated enzyme-producing bacteria that had been isolated from the stinging catfish (*Heteropneustes fossilis*) and the murrel (*Channa punctata*). All tested bacterial strains belonged to the *Bacillus* species. The isolated bacteria could thrive in a wide temperature range, from 20 to 11.0. In their study, among eight isolated bacterial strains, *Bacillus* sp. (HFH4) was found to have the highest cellulolytic activity, followed by *Bacillus* sp. (CPF3) and *Bacillus licheniformis* (CPH6). The highest proteolytic activity was estimated from *B. licheniformis* (CPF4), taken out of murrel. They concluded that the production of proteolytic, cellulolytic, and amylolytic enzymes by bacteria isolated from the gastrointestinal systems of these two Indian air-breathing fish led researchers to hypothesize that these microorganisms are advantageous to the fish's extracellular digestion processes.

The role of two helpful bacteria isolated from the gastrointestinal tract of Atlantic Cod was described by Butt et al. (2021). They studied the role of *Pseudomonas* sp. (GP21) and *Psychrobacter psychrobacter* Sp. (GP12) and

concluded that these two bacteria have an immunomodulatory effect in Cod. They are able to trigger an immunological response in the skin and intestine of cod, which are mucosal surfaces. On the other hand, Caipang et al. (2010) studied about four other bacterial species of occupants of the same fish's intestinal tract. The probiotics *Photobacterium* sp. (GP31), *Shewanella* sp. (GS11), *Psychrobacter* sp. (GP11), and *Vibrio* Sp. were associated with the differential expression of an immune gene connected to bacterial defense and kidney (GV11).

Numerous advantageous outcomes of probiotics have been identified, including the reduction of immune reactions and the prevention of degenerative diseases (Azad et al., 2018). Different routes of application of probiotics in aquaculture and their effects are shown in Fig.1. Different probiotic bacteria, with their source and growth-promoting role, have been presented in Table 1 and Fig. 2.

#### Bacteria-Derived Fertilizers for Aquaculture

For the last two centuries, the aquaculture industry has been growing rapidly and has become a major contributor to global food production. Aquaculture produces a lot of waste, including metabolic byproducts, leftover food, feces, and leftover prophylactic and therapeutic inputs, which degrades water quality and causes disease outbreaks. To improve water quality in aquaculture, a very popular modern approach is the application of beneficial microbes and enzymes to the ponds, known as bioremediation.

According to Wang et al. (2008), to raise the water quality standards in the *Penaeus monodon* culture pond in China *Bacillus* sp. was used, which in turn enhanced the fish's overall growth. Similarly, to improve the water quality, *Bacillus* NL 110 and *Vibrio* sp. NE 17 were added to the *Macrobrachium rosenbergii* culture pond (Mujeeb et al. 2010). Dohail et al. (2009) studied *Lactobacillus acidophilus* growth performance, hematology parameters, and immunoglobulin concentration in African catfish (*Clarias gariepinus*, Burchell 1822) fingerlings. *B. coagulans* SC8168 has the ability to enhance the growth of *Pennaeus vannamei* in aquaculture by improving the water quality parameter (Zhou et al., 2009).

Bacteria are considered to be very good biological transformers. Excess nitrogen application in the aquaculture pond could lead to the accumulation of nitrogenous compounds such as ammonia and nitrite. The waste products of the farmed fish additionally add nitrogenous substances to the pond. By doing nitrification, bacterial microbiota makes ammonia nitrate from ammonia (Kaiser et al., 2013). In the culture pond, two bacterial genera predominantly mediate the process. Nitrosomonas and Nitrobacter both participate in the oxidation of ammonia and nitrite, respectively. By doing this, bacterial species make pond water suitable for fish growth.

Frequent applications of phosphate fertilizer are required to increase fish productivity in the aquaculture pond. It is estimated that about 10% of the fertilizer applied causes an increase in soluble phosphate in the water phase, which is absorbed by the phytoplankton within a few minutes of fertilizer application, whereas the rest is rapidly precipitated, settled at the bottom, and converted into insoluble compounds. (Jana, 2007). Phosphorus is mobilized from organic phosphate esters and inorganic salts, respectively, by enzymes and chelating agents that are extracellular products of the microbial population (organic acids). In a system controlled by humans, the ability of bacteria to dissolve insoluble inorganic phosphate is critical. Numerous phosphate-solubilizing microorganisms, such as bacteria, fungi, and cyanobacteria, can assimilate insoluble inorganic phosphates such as hydroxyapatite, tricalcium phosphate, and rock phosphate and make a significant

**Table 1:** List of different probiotic bacteria with their source and growth-promoting role

Sl. No.	Bacteria	Source Organisms/Medium	Fish growth promoting Role	References
1	<i>Psychrobacter</i> spp.	digestive system of Turbot, wild ( <i>Scophthalmus maximus</i> )	antagonistic activity towards pathogen	(Wanka et al., 2018)
2	<i>Bacillus</i> Sp.	Common carp, <i>Cyprinus carpio</i> ponds	Increase Survival rate Protease activity Lipase activity	(Yanbo and Zirong, 2006)
3	<i>Bacillus</i> spp.	Intestines of <i>Penaeus monodon</i> and shrimp pond sediment	Increase Growth and survival microbial probiotics	Beneficial (Nimrat et al., 2011)
4	<i>L. sakei</i>	Intestines of <i>Paralichthys olivaceus</i>	Macrophage phagocytic activity Peroxidase activity	" (Harikrishnan et al. 2010)
5	<i>Escherichia coli</i> , <i>Proteus vulgaris</i> , <i>Bacillus subtilis</i> , <i>Klebsiella pneumoniae</i> , <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i>	<i>Scomberomorus guttatus</i>	-	(Karthiga et al., 2016)
6	<i>Bacillus</i> Sp. (HFH4, CPF3), <i>Bacillus licheniformis</i> (CPH6, CPF4)	The gastrointestinal tract of <i>Channa punctatus</i> and <i>Heteropneustes fossilis</i>	Proteolytic, Cellulolytic, and amylolytic and enzyme activity	(Banerjee et al., 2013)
7	<i>Pseudomonas</i> Sp. (GP21) and <i>psychrobacter</i> Sp. (GP12)	The gastrointestinal tract of Atlantic Cod	Elicit the immune response, particularly in mucosal surfaces such as the skin and intestine.	(Butt, et al., 2021)
8	<i>Psychrobacter</i> Sp. (GP11), <i>Shewanella</i> Sp. (GS11), <i>Photobacterium</i> Sp. (GP31), and <i>Vibrio</i> Sp. (GV11),	Atlantic cod's intestine	Differential expression of an immune gene associated with bacterial defense and inflammation in the head kidney leukocytes.	(Caipang et al., 2010)
9	<i>Nitrosomonas Nitrobacter</i> Sp.	The gut of Catla ( <i>Catla catla</i> ), Rohu ( <i>Labeo rohita</i> ), and Grass carp ( <i>Ctenopharyngodon idella</i> )	Block the pathogen entry in the culture pond and can raise the concentration of dissolved oxygen.	(Sunitha and Krishna, 2016)
10	<i>Bacillus</i> Sp.,	Added in rearing water	Increasing growth through improving the water quality in the culture pond	(Wang et al., 2008)
11	<i>Bacillus</i> NL 110, <i>Vibrio</i> Sp. NE 17	Added in rearing water	Enhancing water quality in the culture pond	(Mujeeb et al., 2010)
12	<i>Lactobacillus acidophilus</i>	Added in rearing water	Enhancing water quality in the culture pond	(Dohail et al., 2009)
13	<i>B. coagulans</i> SC8168	Added in rearing water	Enhancing water quality in the culture pond	(Zhou et al., 2009)
14	<i>Nitrosomonas Nitrobacter</i>	Applied in culture pond	Promote growth by Oxidizing ammonia and nitrite in the cultured pond.	(Kaiser et al., 2013)
15	<i>Bacillus</i> spp.	Added in the surrounding water	Increase growth performance in cultured pond by phosphate solubilization and phosphate production	(Jana, 2007)
16	<i>Methylococcus capsulatus</i>	Added to the diet of Atlantic salmon ( <i>Salmo salar</i> )	Enhance growth by supplying abundant protein to <i>Salmo salar</i>	(Romarheim et al., 2011)
17	<i>Clostridium butyricum</i>	kuruma shrimp ( <i>Marsupenaeus japonicas</i> )	Increased the content of intestinal short-chain fatty acid along with propionic acid and butyric acid, and increased the levels of body crude protein.	(Duan et al., 2018)
18	Yeast <i>Debaryomyces hansenii</i> HF1	Supplemented to Sea bass larvae	Promote growth by secreting amylase and trypsin, enzymes that help in digestion	(Tovar et al., 2002)
19	<i>Bacillus cereus</i>	Added to the meal of Juvenile <i>Dentex dentex</i> L	Increases fish growth due to a greater food consumption capacity	(Merrifield et al., 2010)
20	<i>B. subtilis</i> , <i>B. licheniformis</i> , and <i>Enterococcus faecium</i>	Supplemented with a traditional meal of Rainbow trout	Enhance fish growth by increasing feed consumption efficiency	(Merrifield et al., 2010)
21	<i>Bacillus subtilis</i>	Gut of <i>Cirrhinus mrigala</i>	Increase the length and weight of guppies swordtail ( <i>Xiphophorus helleri</i> , <i>X. maculatus</i> ), and increase the proteases and amylases activity in ornamental fish	(Ghosh et al., 2008)
22	<i>Carnobacterium</i> Sp.	Supplemented to salmon bowel	Promote growth by showing <i>in vitro</i> antagonism against known fish pathogens in rainbow trout and Atlantic salmon	(Robertson et al., 2000; Martinez et al., 2012)
23	<i>Bacillus cereus</i> , <i>Paenibacillus polymyxa</i> , and <i>Pseudomonas</i> Sp. PS-102	-	Act as the antagonist against various pathogens in shrimp	(Ravi et al., 2007; Vijayan et al., 2006)
24	<i>Lactobacillus rhamnosus</i>	Administrated to <i>Oreochromis niloticus</i>	Stimulate the phagocytic activity in Tilapia	(Cano-Lozano et al., 2022)
25	<i>Lactobacillus Plantarum</i> and <i>Pseudomonas fluorescent</i>	Added in pond water	Increase survival rate by 96.22% in <i>Clarias gariepinus</i>	(Omenwa et al., 2015)
26	<i>Lactobacillus acidophilus</i> (Fla) and <i>Saccharomyces boulardii</i> (FSB)	Supplemented with the diet of <i>Acipenser baerii</i>	Enhance growth, immunity, and survival rate	(Mocanu et al., 2022)

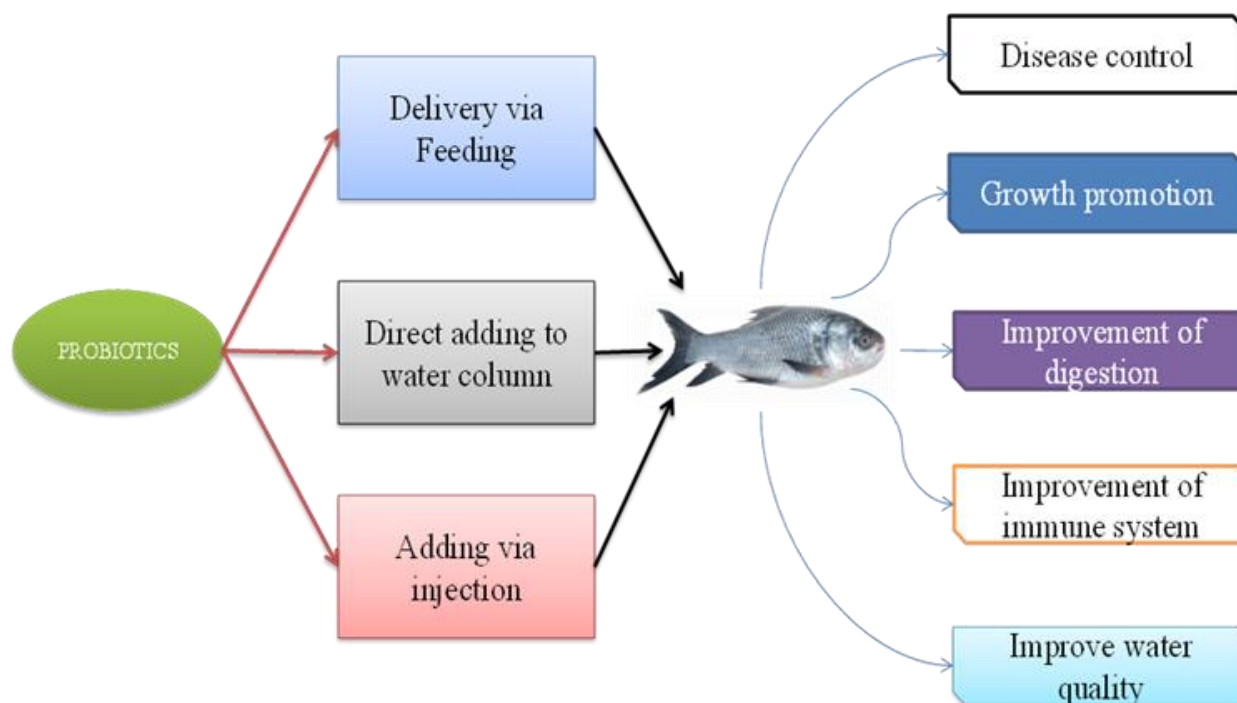


Fig. 1: Different routes of application of probiotics in aquaculture and their effect.

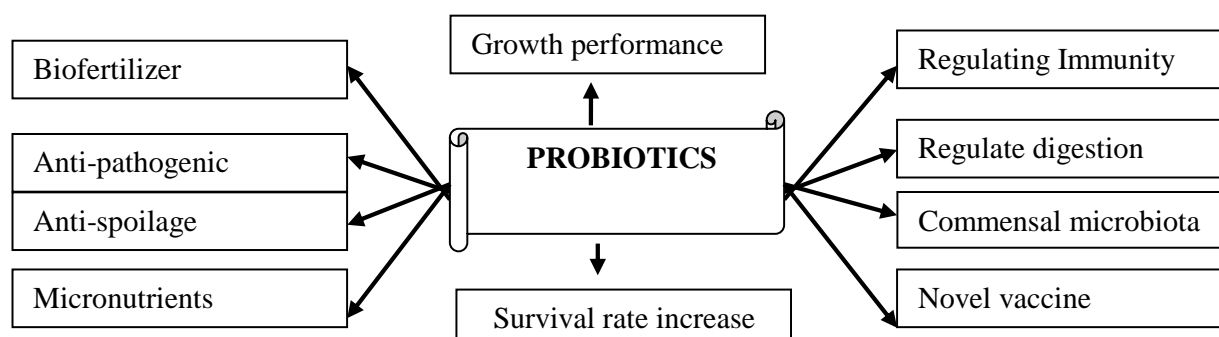


Fig. 2: Role of Fish Growth Promoting Bacteria.

portion of them soluble by producing organic and inorganic acids in the water and sediments of fishponds. *Bacillus* is the bacteria that produces the most phosphates and is the most effective at solubilizing phosphates, according to reports (Jana, 2007).

#### Bacteria as an Alternative Feed for Aquaculture

Fish are a good source of protein as well as vitamins, fatty acids, and minerals for human health. Aquaculture industries were developed to increase fish production as per the requirements of human health. For proper growth, strong immunity, stress tolerance, and good reproductive ability in the aquaculture industries, fish require an abundant proteinaceous diet rather than traditional fish meals. A wide range of bacteria rich in protein can make up 65% of total dry weight (Wang et al., 2020). Gamboa Delgado and Marquez (2018) that dietary supplementations of beneficial bacteria can be used for better growth, immunity, and stress response in aquatic animals and can also improve diet palatability.

It was discovered that methanotrophic bacteria, particularly *Methylococcus capsulatus*, when employed as a replacement for traditional fish meal in the case of Atlantic salmon (*Salmo salar*), could be a substitute protein source for fish meal. According to Perera et al. (1995), when the traditional fish meal was replaced by the bacterial single-cell

protein, it was noted that the growth rate, amount of feed consumed, and rate of absorption of rainbow trout were unchanged (*Oncorhynchus mykiss*). When beneficial bacteria were used as feed, the growth rate of black tiger prawns significantly changed from lower to higher (Glencross et al., 2014). In addition, when bacterial protein was added to the shrimp feed, improvements in growth were recorded (Arnold et al. 2016). According to De Schryver et al. (2008), probiotic bacteria have the capacity to enhance rearing water quality parameters by forming some flocculated materials, resulting in a higher breeding density in aquaculture. According to Dantas et al. (2016), when 20% of a fish diet is replaced with a prepared food, it may actually boost shrimp growth. A similar result was recorded by Chen et al. (2018) and Kim et al. (2018) in the case of flatfish (*Paralichthys olivaceus*) as well as sea cucumber (*Apostichopus japonicas*).

According to Perera et al. (1995), overuse of bacterial biomass showed a negative feedback mechanism, resulting in reduced growth and low digestibility of nutrients caused by the decrease in nitrogen (N) absorption and increase in urea excretion. According to Goodall et al. (2016), although domesticated broodstock's reproductive efficiency was unaffected, a higher amount of bacterial biomass highly affected the egg-hatching rate of *Penaeus monodon*.

### Bacteria Generate Beneficial Micronutrients

Aquatic animals fulfill their basic nutrient requirements through feed intake. But micronutrients such as vitamins, fatty acids, and some essential amino acids that promote growth and normal physiological functions may not be sufficiently available in the feed provided (Fig. 2). Therefore, it is needed to supply it artificially. Recent studies have reported that a number of beneficial bacteria can produce some essential micronutrients. So probiotic-supplemented diets are now becoming a famous concept for aquaculture industries.

Cyanocobalamine, commonly known as vitamin B-12, is one of the essential cofactors for DNA synthesis and also plays a role in the metabolism of fats and amino acids, which are generated by some probiotic strains (Wang et al., 2020). Teshima and Kashiwada (1967) isolated some bacterial strains from the intestine of carp and demonstrated that these strains are capable of producing B-12 vitamins in a large percentage. Even in the absence of dietary B-12, these helpful bacteria may provide vitamin B-12 for the typical growth of aquatic species (Wang et al., 2020). Another report stated that the intestinal microbiota of channel catfish creates roughly 14 nanograms of cyanocobalamin per gram of body weight, which the host organism absorbs directly into its d. Because of this, a dietary supplement of B-12 is not necessary (Wang et al., 2020). According to LeBlanc et al. (2013) and (2017), various vitamins from the B family and vitamin K that are produced by the gut bacteria of humans have a similar impact on the host body.

However, plant protein, a substitute for fish meals, may also be deficient in several necessary amino acids, such as lysine, tryptophan, and sulfur-containing amino acids. As a result, shrimp development performance may be affected when using plant protein sources. To address this issue, Jannathulla et al. (2017) and associates employed bacterial, fungal, and yeast fermentation processes. They then measured the increase in critical amino acids, as expected. Trypsin inhibitors, phytic acid, saponin, tannin, glucosinolate, and guar gum, which are antinutrients found in plant protein sources, were also reduced after the fermentation process. Jannathulla et al. (2017) concluded from their study that plant protein sources fermented with beneficial microorganisms yield a higher growth ratio when fed to the aquatic fauna.

Probiotics create simple and short-chain fatty acids by fermenting some specific intestine-found fiber, according to LeBlanc et al. (2017). They also play a role in the host's energy metabolism. Duan et al. (2018) demonstrated that the probiotic *Clostridium butyricum*, when supplemented with the diet of *Marsupenaeus japonicas*, enhanced the short-chain fatty acid content of the intestine along with butyric acid and propionic acid, which can increase body crude protein and encourage kuruma shrimp growth. According to Hao et al. (2017), this kind of diet can alter the grass carp's hindgut microbiota, which is related to the concentrations of short-chain fatty acids.

### Bacteria Regulate Digestion in the Host

According to Balcázar et al. (2006), beneficial microorganisms show a good effect on the metabolic process of cultured animals in aquaculture by synthesizing extracellular enzymes such as proteases, lipases, and amylases, along with providing some micronutrients like vitamins, amino acids, and fatty acids. Therefore, when probiotics are added to the diet, the nutrients are absorbed by the host organisms more effectively (El-Haroun et al., 2006). Probiotics can therefore be used to improve how well edible fish digest. According to a study by Tovar et al. (2002), *Debaryomyces hansenii* HF1, which is a yeast

species, can produce the polyamines spermine and spermidine, which are crucial for the development of the mammalian gastrointestinal tract. Additionally, this yeast secretes trypsin and amylase, which help sea bass larvae digest their food. It has been reported that when the diet of juvenile *Dentex dentex* L. is supplemented with 0.5 g of *Bacillus cereus* strain Ekg<sup>-1</sup> of food, it increases fish growth due to a greater food consumption capacity (Merrifield et al., 2010). When the diet of rainbow trout was supplemented with *B. subtilis*, *B. licheniformis*, and *Enterococcus faecium* for 10 weeks, similar results were found (Merrifield et al., 2010).

When diets of *Litopenaeus vannamei* (white shrimp) Boone and *Fenneropenaeus indicus* are treated with 50g of probiotics per kg of meal, the digestion of dry matter, crude protein, and phosphorus rises along with the larger body size. In order to create a consistent impact on the synthesis of enzymes involved in digestion, probiotic maintenance in all ontogenetic phases of shrimp has been explicitly highlighted in other studies (Martinez et al., 2012). When *B. subtilis* was taken out of the digestive tract of *C. mrigala* and supplemented with the diet of guppies (*Poecilia reticulata*, *P. sphenops*) and swordtails (*Xiphophorus helleri*, *X. maculatus*), these fish displayed a larger body weight and good length, along with the remarkable functions of enzyme proteases and amylases of the gut (Ghosh et al., 2008). A wide range of coenzymes is secreted by *Bacillus*, which in turn increases the enzymatic digestion of the fish bodies. Protease, cellulase, lipase, chitinase, and trypsin activity have also been demonstrated by these isolated beneficial bacteria from the digestive tract of aquatic animals (Vine et al., 2006).

### Beneficial Bacteria have an Anti-Pathogen Action

In the past centuries, antibiotics have been used as potent anti-pathogenic drugs in aquaculture. Later, it was found that though antibiotics can prevent disease in aquatic crops, they have some negative impacts on cultured aquatic animals as well as on human health. Problems raised by the applications of antibiotics in aquaculture industries include an accumulation of antimicrobial drug residue in host tissue, the development of microbial defiant mechanisms, and the disbalance of beneficial gut fauna in cultured organisms, which further affects their physique (Nakano, 2007). Even antibiotic use in organisms intended for human consumption is governed by EU regulations. (Ronson and Medina, 2002). People are concerned about their health, and it has been seen that natural items without additions like antibiotics are always preferred. In addition, disease prevention is more common than disease treatment. Therefore, using probiotics to inhibit pathogens and reduce disease in aquaculture species is a viable alternative. (Martinez et al., 2012).

Beneficial bacteria can stimulate innate and humoral immunity in fish to combat pathogenic microorganisms (Allameh et al., 2017). By secreting some chemical substances, probiotic bacteria can prevent the colonization by pathogens within the gut of the host organisms. The creation of antibiotics, bacteriocins, siderophores, digestive enzymes like lysozymes, proteases, and hydrogen peroxide, as well as the adjustment of the gut pH brought on by the formation of organic acids, are what give probiotics their antibacterial effects. (Verschuere et al., 2000).

Taoka and Maeda Jo (2006) demonstrated that live beneficial microorganism administration to tilapia, *Oreochromis niloticus*, enhanced innate immune response, which can be measured by lysozyme activity, neutrophil migration, and bactericidal activity, responsible for the improvement of the defense mechanism of fish against the

infection caused by *Edwardsiella tarda*. Live *Carnobacterium* spp. were given to rainbow trout and Atlantic salmon by Robertson et al. (2000) and isolated from the salmon bowel, which demonstrated in vitro antagonistic activity against recognized fish pathogens like *Aeromonas hydrophila*, *A. salmonicida*, *Flavobacterium psychrophilum*, *Photobacterium damsela*, and *Vibrio* spp. There is proof that deceased probiotic cultures made up of a combination of *Aeromonas hydrophila* A3-51, *Carnobacterium* BA211, and *Vibrio fluvialis* A3-47S can help control furunculosis disease in rainbow trout (Martinez et al., 2012). Probiotics, including *Bacillus cereus*, *Paenibacillus polymyxa*, and *Pseudomonas* spp., have been evaluated in studies. In the case of shrimp, PS-102 biocontrol drugs prevent a variety of *Vibrio* species (Ravi et al., 2007; Vijayan et al., 2006).

It has been observed that microbial strains that were taken out from the gastrointestinal mucosa of clownfish have the ability to generate antimicrobial metabolites in vivo conditions which were useful to deactivate some pathogenic bacteria such as *A. hydrophila* and *Vibrio alginolyticus* among others (Vine et al., 2004). According to Ghosh et al. (2008) probiotic bacteria with concentrations of  $10^6$  to  $10^8$  cells $g^{-1}$  enhanced the proliferation of beneficial microorganisms in the GI tract of ornamental fishes which belong to the genera *Poecilia* and *Xiphophorus*.

### Regulation of Fish Immune Systems by Beneficial Microorganisms

Probiotics are now a crucial component of aquaculture methods that can help achieve high productivity. Commonly used beneficial microbes in aquaculture include *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Enterococcus*, *Carnobacterium*, *Shewanella*, *Bacillus*, *Aeromonas*, *Vibrio*, *Enterobacter*, *Pseudomonas*, *Clostridium*, and *Saccharomyces* species (Fernandes et al., 2021). Probiotics have been shown to have a variety of positive effects on fish, including stimulation of growth, improved nutrition, disease resistance, and other positive effects. But one of the most frequently cited effects is the modification of the immune system in aquatic organisms. Probiotic supplementation can elevate phagocytic, lysozyme, complement, and respiratory burst activity as well as the expression of various cytokines in fish. According to studies, microorganisms can enhance the number of immunoglobulin cells and acidophilic granulocytes in the fish gut immune system (Fernandes et al., 2021).

Pirarat et al. (2006) recorded stimulation of phagocytic activity in *Oreochromis niloticus* (Tilapia) when administered with *Lactobacillus rhamnosus* for 2 weeks. The phagocytic activity of *O. mykiss* has also been shown to increase when *Clostridium butyricum* is fed to tilapia (Sharifuzzaman and Austin, 2009). *Bacillus subtilis* and some members of the LAB group have been shown to enhance respiratory burst activity in fish (Nikoskelainen et al., 2003; Salinas et al., 2005; Salinas et al., 2006; Zhou et al., 2009). Bacteria like *Lactobacillus rhamnosus*, *Carnobacterium maltaromaticum*, and *Carnobacterium divergens* enhance the lysozyme level in *O. mykiss* (Kim and Austin, 2006; Panigrahi et al., 2004). A study has shown that when *Ulva* Sp. was fed to Nile tilapia, it significantly enhanced the complement activity (i.e., alternative hemolytic complement activity (ACH<sub>50</sub>), an essential component of the innate immunity system), while the muscle colour was modified, and the sour parameter of the fillets was decreased (Valente et al., 2016). *Pseudomonas aeruginosa*, according to Kanther et al. (2011), can also affect the immune system of larval zebrafish by promoting NF- $\kappa$ B-dependent production of innate immunity genes, including complement factor b (cfb) and serum amyloid a (saa), which increase neutrophil

infiltration. A study reveals that a unique protein, AimA, is secreted by *Aeromonas veronii* in the zebra fish intestine and is beneficial for both the host and microbes. It protects the host by preventing chemical and bacterially induced inflammation, and it protects *A. veronii* by helping in colonization and the host immune response (Rolig et al., 2018). According to Panigrahi et al. (2007), microbes like *L. rhamnosus*, *E. faecium*, and *B. subtilis* can increase the level of pro-inflammatory cytokines like IL-1b and TGF $\beta$  in Tilapia's head, kidney, and spleen. Likewise, the anti-inflammatory response in Tilapia is also influenced by *C. maltaromaticum* and *C. divergens*, which can express IL-1b, IL-8, TNF-a, and TGF-b in the head and kidney of *O. mykiss* (Kim and Austin, 2006). Picchiatti et al. (2009) recorded the enhancement of T-lymphocytes in the GI tract of *D. labra* when supplemented with *L. delbrueckii* sp. through artemia and rotifers.

### Commensal Microbiota in Host Immune Homeostasis Maintenance

Healthy fish have various microbial communities in contrast to those that are ill, which provides clear evidence of the intimate relationship between commensal microbiota and bodily homeostasis. This remark is well demonstrated by the comparison of a healthy member of the same species with a crucian carp (*Carassius auratus*) that has red operculum disease and a varied microbial population. On the other hand, some species of microbes are found abundantly in diseased fish as good pathogens, and these include *Vibrio*, *Aeromonas*, and *Shewanella* (Li et al., 2017). The same record was found in the case of *Coreius guichenoti* when infected by the bacterial pathogen *Aeromonas salmonicida*, resulting in furunculosis (Li et al., 2016). According to Nie et al. (2017), when *Plecoglossus altivelis* was infected by *V. anguillarum*, it was demonstrated that the host's gut microbiota dynamics significantly decreased, along with the complexity, connectivity, and cooperativeness of gut bacterial interspecies interaction. This affects the host immunological response, which includes TNF $\alpha$  and IL-1b expression. According to the findings of this study, the host defense system and the inhabitants of the gut interact to prevent harmful microbes.

When antibiotics were used to treat various pathogenic microorganisms in the host, it was seen that they affected the commensal microbiota in the host intestine, which further affected the host immune response. Moreover, Limbu et al. (2018) observed human health risks. To overcome this problem, He et al. (2017) used olaquinox on zebrafish against the pathogen *A. hydrophila* infection, and they found a compromised result, i.e., they recorded minimal changes in the intestinal microbiota and host immune response of zebrafish. They came to the conclusion that bacteria may have an impact on the host immune system.

### Beneficial Bacteria Modulate Commensal Microbiota

The gut microbiota of fish is undoubtedly abundant and diverse and is capable of providing a potential barrier for foreign invaders such as pathogenic organisms like viruses, bacteria, protozoans, parasites, nematodes, etc. These intestinal microbiotas maintain a mutual relationship with the host for nutrition, defense, hormonal, neural, and metabolic activity. An imbalance in the intestinal microbiota of Pisces causes critical health conditions resulting in growth retardation, low immunity, various clinical conditions, and a higher mortality rate for the host. Several therapeutic strategies are applicable to overcoming this problem, and one of the most modern is the administration of probiotic bacteria to the fish.

Fish that have been treated with antibiotics may have a decline in their gut microbiota, but probiotics can help restore the balance (Yukgehnai et al., 2020). Probiotics, as stated by Reid et al. (2003), can influence the gut microbiota through mechanisms such as colonization of the intestine mucosa, competitive adherence and exclusion, and the production of beneficial substances and chemical signs for the host, which in turn influence the immune system to modulate and regulate allergic responses. Pathogen growth can be inhibited by the presence of chemicals in probiotics such as bacteriocins, enzymes, hydrogen peroxide, and organic acids (Alonso et al., 2019). Tan et al. (2019) report that in the intestine of Tilapia, *R. stabekisii* stimulates the growth of other probiotic bacteria, specifically *Bacillus* and *Lactobacillus spp.*, while concurrently decreasing the number of harmful bacteria, such as *Streptococcus* and *Staphylococcus spp.* According to research published by Huyben et al. (2017), the yeast *Saccharomyces cerevisiae* can alter the composition of the gut microbiota of fish without causing any harm to the host. Yeast protein is a dietary supplement that has been shown to promote beneficial yeast species in fish. According to research by Huyben et al. (2018), supplementing a rainbow trout's diet with live yeast has been shown to increase the diversity of yeast species in the fish's digestive tract. When immersed in a solution containing 49 different strains of *Lactobacillus*, He et al. (2017) found that the zebrafish's intestines became gradually colonized and that the *Lactobacillus* was distributed dynamically along the length of the intestine.

#### Increase Survival Rate

Probiotics are live beneficial microorganisms that are capable of maintaining good health, desirable growth, strong immunity, and a higher survival rate in aquatic organisms in aquaculture practice. Probiotic supplementation with the fish diet for obtaining desirable growth and a higher survival rate is a famous approach in aquaculture. Definite probiotic doses in fish can enhance the gut microflora, inhibit pathogens, and increase immunity in the host.

Omenwa et al. (2015) also reported that the addition of *Lactobacillus plantarum* and *Pseudomonas fluorescens* in the pond water of *C. gariepinus* juveniles increases the survival rate by 96.22%. Likewise, Tarnecki et al. (2019) supplemented a mixture of *Bacillus* species (*B. licheniformis* and *B. amyloliquefaciens*) to the larval common snook (*Centropomus undecimalis*) in three different ways: 1) probiotic added to rearing water and live feed; 2) probiotic supplemented with rearing water only and 3) no probiotic control. After some days, they recorded the results of each treatment. From these two separate trial treatments, they discovered a 2.5-fold increase in snook survival after adding the probiotic. And a 20% increase in survival seven days after a transit event. Mocanu et al. (2022) showed how probiotics can increase the survival rate of aquatic organisms in a rearing pond. They carried out an experiment on Siberian sturgeon (*Acipenser baerii*) in a recirculating system. The diet of *A. baerii* is supplemented with *Saccharomyces boulardii* (FSB) and *L. acidophilus* (Fla) separately as well as combined. They feed 2000 fish with four different diets for 8 weeks: 1) a diet without probiotic addition; 2) a diet with *Lactobacillus acidophilus*; 3) a diet with *Saccharomyces boulardii* and 4) a diet with a mixture of both bacteria and yeast in equal proportion. The Fla+FSB diet results in higher growth performance, a higher feed conversion ratio, strong immunity, and higher survival than the control diet in the *A.r. baerii*.

#### Application of Probiotics in Biofloc Technology

Biofloc technology is a fish farming system that recycles waste nutrients as fish food. By manipulating the water's carbon-nitrogen ratio, heterotrophic bacteria can be promoted to proliferate and utilized in situ by cultured animals or extracted and processed as feed ingredients.

#### Novel Vaccine

Another innovative application is the diverse synthesis of functional genes in bacteria and their application in aquaculture activities. Jia and colleagues engineered the heterocystous cyanobacterium *Anabaena Sp.* PCC 7120 to express the principal envelope protein (VP28) of the white spot syndrome virus for oral immunization (Jia et al., 2016).

As previously mentioned, many bacterial strains have been demonstrated to be favorable to both aquatic creatures and humans. There is no information on whether an aquaculture strain will directly or indirectly enhance consumer health through aquatic products. Contrarily, there are currently relatively few practical applications despite the fact that numerous studies have shown that different bacterial strains have a positive impact on aquaculture growth. As a result, there are many opportunities for beneficial bacteria to increase the quantity and quality of aquaculture and to provide wholesome aquatic products. For this reason, the corresponding basic research and commercial trials are important.

#### Future Direction-Novel Applications of Beneficial Bacteria

The good effects of beneficial bacteria on aquatic organisms have led to significant research and widespread application of these microbes in aquaculture, as well as the development of several novel applications. The nutritional and healthcare applications of these interventions stand to benefit from the novel types, mechanisms, and applications that are currently under investigation. Related domains of microbiome-targeted therapies are growing, and the environment for implementation across regulatory, policy, prescriber, and consumer sectors is shifting, heralding a new era of profound transformation. Widespread improvements in data gathering and analytical methods allow for the discovery of novel candidates for probiotics and prebiotics and provide light on the complex relationships between these microbes and the hosts they inhabit. Increased focus is being placed on discovering novel uses for probiotics and prebiotics in a wide variety of medical contexts, anatomical locations, demographic groupings, and administration methods. The incorporation of probiotics and prebiotics into nutrition and healthcare is also influenced by the development of regulatory frameworks, clinical guidelines, and market tendencies. Future healthcare solutions may include the use of probiotics and prebiotics to combat microbial infections and other conditions. In order to treat and prevent COVID-19 infection, probiotics and prebiotics have been proposed as potential components of both preventative and acute care methods (Villena and Kitazawa, 2020).

#### Conclusion

The aquaculture business has experienced a remarkable expansion in recent years because of the rising demand for seafood and the advancement of cultural technologies. However, the contemporary aquaculture business, particularly high-density aquaculture, has a number of significant obstacles. These are the lack of a cheap and plentiful source of protein for feeding, the significant loss brought on by viral infection, and the quality loss during growth and storage. Bacteria can be employed as a potential remedy for these issues due to their

inexpensive cost, high protein content, and variety of biological activities. Numerous beneficial bacterial species have been studied and utilized in aquaculture for a variety of purposes. However, fresh applications and pertinent mechanistic research will be required in the future to improve current procedures and avoid potential hazards.

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#### Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

#### REFERENCES

- Allameh, S.K., Noaman, V. and Nahavandi, R. (2017). Effects of probiotic bacteria on fish performance. *Adv Tech in Clin Microbiol* 1(2):11: <http://www.imedpub.com/advanced-techniques-in-clinicalmicrobiology>
- Alonso, S., Castro, M.C., Berdasco, M., de la Banda, I.G., Moreno-Ventas, X. and de Rojas, A.H. (2019). Isolation and partial characterization of lactic acid bacteria from the gut microbiota of marine fishes for potential application as probiotics in aquaculture. *Probiotics Antimicrob Proteins* 11:569-579 <https://doi.org/10.1007/s12602-018-9439-2>
- Arnold, S., Smullen, R., Briggs, M., West, M. and Glencross, B. (2016). The combined effect of feed frequency and ration size of diets with and without microbial biomass on the growth and feed conversion of juvenile *Penaeus monodon*. *AquacNutri* 22(6):1340-1347. <https://doi.org/10.1111/anu.12338>
- Azad, M., Kalam, A., Sarker, M., Li, T. and Yin, J. (2018). Probiotic species in the modulation of gut microbiota: an overview. *BioMed Research International* 2018:1-8 <https://doi.org/10.1155/2018/9478630>
- Balcázar, J.L., De Blas, I., Ruiz-Zarzuola, I., Cunningham, D., Vendrell, D. and Múzquiz, J.L. (2006) The role of probiotics in aquaculture. *Vet Microbiol* 114(3-4):173-186. <https://doi.org/10.1016/j.vetmic.2006.01.009>
- Banerjee, G., Ray, A.K. and Askarian, F. (2013). Characterization and identification of enzyme-producing autochthonous bacteria from the gastrointestinal tract of two Indian air-breathing fish. *Benef Microbes* 4(3):277-284. <https://doi.org/10.3920/BM2012.0051>
- Butt, U. D., Lin, N., Akhter, N., Siddiqui, T., Li, S. and Wu, B. (2021). Overview of the latest developments in the role of probiotics, prebiotics, and synbiotics in shrimp aquaculture. *Fish & Shellfish Immunology* 114, 263–281. <https://doi.org/10.1016/j.fsi.2021.05.003>
- Caipang, C.M.A., Brinchmann, M.F. and Kiron, V. (2010). Antagonistic activity of bacterial isolates from intestinal microbiota of Atlantic cod, *Gadus morhua* and an investigation of their immunomodulatory capabilities. *Aquac Research* 41(2):249-256. <https://doi.org/10.1111/j.1365-2109.2009.02327.x>
- Cano-Lozano, J. A., Villamil Diaz, L. M., Melo Bolivar, J. F., Hume, M. E. and Ruiz Pardo, R. Y. (2022). Probiotics in tilapia (*Oreochromis niloticus*) culture: Potential probiotic *Lactococcus lactis* culture conditions. *Journal of Bioscience and Bioengineering* 133(3), 187–194. <https://doi.org/10.1016/j.jbiosc.2021.11.004>
- Chen, J., Liu, P., Li, Y., Li, M. and Xia, B. (2018). Effects of dietary biofloc on growth, digestibility, protein turnover and energy budget of sea cucumber *Apostichopus japonicus* (Selenka). *Anim Feed Sci Technol* 241:151-162. <https://doi.org/10.1016/j.anifeedsci.2018.05.002>
- Dailey, F. E., Turse, E. P., Rossow, B., Kuwajima, V. K. and Tahan, V. (2019). Probiotics for Gastrointestinal and Liver Diseases: An Updated Review of the Published Literature. *Endocrine, Metabolic & Immune Disorders Drug Targets* 19(5), 549–570. <https://doi.org/10.2174/1871530318666181022163944>
- Dantas, Jr E.M., Valle, B.C.S., Brito, C.M.S., Calazans, N.K.F., Peixoto, S.R.M. and Soares, R.B. (2016). Partial replacement of fishmeal with biofloc meal in the diet of postlarvae of the Pacific white shrimp *Litopenaeus vannamei*. *Aquac nutri* 22(2):335-342. <https://doi.org/10.1111/anu.12249>
- De Schryver, P., Crab, R., Defoirdt, T., Boon, N. and Verstraete, W. (2008). The basics of bio-flocs technology: the added value for aquaculture. *Aquac* 277(3-4):125-137. <https://doi.org/10.1016/j.aquacultur.e.2008.02.019>
- Dohail, A., Abdullah, M., Roshada, H. and Aliyu, M. (2009). Effects of the probiotic, *Lactobacillus acidophilus*, on the growth performance, hematology parameters and immunoglobulin concentration in African Catfish (*Clarias gariepinus*, Burchell 1822) fingerling. *Aquac Research* 40(14):1642-1652. <https://doi.org/10.1111/j.1365-2109.2009.02265.x>
- Duan, Y., Dong, H., Wang, Y., Zhang, Y. and Zhang, J. (2018). Effects of the dietary probiotic *Clostridium butyricum* on intestine digestive and metabolic capacities, SCFA content and body composition in *Marsupenaeus japonicus*. *Journal Ocean University China* 17(3):690-696 <https://doi.org/10.1007/s11802-018-3464-3>
- El-Haroun, E.R., Goda, A.S. and Chowdhury, M.A. (2006). Effect of dietary probiotic Biogen supplementation as a growth promoter on growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (L). *Aquac Research* 37(14):1473-1480. <https://doi.org/10.1111/j.1365-2109.2006.01584.x>
- El-Saadony, M.T., Alagawany, M., Patra, A.K., Kar, I., Tiwari, R., Dawood, M.A.O., Dhama, K. and Abdel-Latif, H.M.R. (2021). The functionality of probiotics in aquaculture: An overview. *Fish Shellfish Immunology* Oct;117:36-52. <https://doi.org/10.1016/j.fsi.2021.07.007> PMID: 34274422.
- FAO (2020). The State of World Fisheries and Aquaculture 2020. Sustainability in action. Rome.
- FAO (2022). The State of World Fisheries and Aquaculture 2022. Towards Blue Transformation. Rome, FAO. <https://doi.org/10.4060/cc0461en>
- Fernandes, S., Kerkar, S., D'Costa, A., Costa, M., Mishra, A., Shyama, S. K. and Das, K. R. (2021). Immunostimulatory effect and toxicology studies of salt pan bacteria as probiotics to combat shrimp diseases in aquaculture. *Fish & Shellfish Immunology* 113, 69–78. <https://doi.org/10.1016/j.fsi.2021.03.017>
- Gamboa Delgado, J. and Marquez Reyes, J.M. (2018). Potential of microbial derived nutrients for aquaculture development. *Review Aquac* 10(1):224-246. <https://doi.org/10.1111/raq.12157>
- Ghosh, S., Sinha, A. and Sahu, C. (2008). Dietary probiotic supplementation on growth and health of live-bearing ornamental fishes. *Aquac Nutrition* 14(4):289-299. <https://doi.org/10.1111/j.1365-2095.2007.00529.x>
- Glencross, B., Irvin, S., Arnold, S., Blyth, D., Bourne, N. and Preston, N. (2014). Effective use of microbial biomass products to facilitate the complete replacement of



- fishery resources in diets for the black tiger shrimp, *Penaeus monodon*. *Aquac* 431:12-19. <https://doi.org/10.1016/j.aquaculture.2014.02.033>
- Goodall, J.D., Wade, N.M., Merritt, D.J., Sellars, M.J., Salee, K. and Coman, G.J. (2016). The effects of adding microbial biomass to grow-out and maturation feeds on the reproductive performance of black tiger shrimp, *Penaeus monodon*. *Aquac* 450:206-212. <https://doi.org/10.1016/j.aquaculture.2015.07.036>
- Govindaraj, K., Samayanpaulraj, V., Narayanadoss, V. and Uthandakalaipandian, R. (2021). Isolation of Lactic Acid Bacteria from Intestine of Freshwater Fishes and Elucidation of Probiotic Potential for Aquaculture Application. *Probiotics Antimicrob* 13(6):1598-1610.
- Hao, Y.T., Wu, S.G., Jakovli, I., Zou, H., Li, W.X. and Wang, G.T. (2017). Impacts of diet on hindgut microbiota and short chain fatty acids in grass carp (*Ctenopharyngodon idellus*). *Aquac Research* 48(11): 5595-5605. <https://doi.org/10.1111/are.13381>
- Harikrishnan, R., Balasundaram, C. and Heo, M.S. (2010). *Lactobacillus sakei* BK19 enriched diet enhances the immunity status and disease resistance to streptococcosis infection in kelp grouper, *Epinephelus bruneus*. *Fish Shellfish Immunology* 29(6):1037-1043. <https://doi.org/10.1016/j.fsi.2010.08.017>
- He, S., Ran, C., Qin, C., Li, S., Zhang, H., De Vos, W.M., Ringø, E. and Zhou, Z. (2017) Anti- infective effect of adhesive probiotic *Lactobacillus* in fish is correlated with their spatial distribution in the intestinal tissue. *Science Rep* 7:1-12. <https://doi.org/10.1038/s41598-017-13466-1>
- He, S., Wang, Q., Li, S., Ran, C., Guo, X., Zhang, Z. and Zhou, Z. (2017). Antibiotic growth promoter olaquinox increases pathogen susceptibility in fish by inducing gut microbiota dysbiosis. *Science China-Life Science* 60:1260-1270. <https://doi.org/10.1007/s11427-016-9072-6>
- Huyben, D., Nyman, A., Vidaković, A., Passoth, V., Moccia, R., Kiessling, A., Dicksved, J. and Lundh, T. (2017) Effects of dietary inclusion of the yeasts *Saccharomyces cerevisiae* and *Wickerhamomyces anomalus* on gut microbiota of rainbow trout. *Aquac* 473:528-537. <https://doi.org/10.1016/j.aquaculture.2017.03.024>
- Huyben, D., Sun, L., Moccia, R., Kiessling, A., Dicksved, J. and Lundh, T. (2018). Dietary live yeast and increased water temperature influence the gut microbiota of rainbow trout. *Journal Appl Microbiology* 124:1377-1392. <https://doi.org/10.1111/jam.13738>
- Jana, B.B. (2007). Distribution pattern and role of phosphate solubilizing bacteria in the enhancement of fertilizer value of rock phosphate in aquaculture ponds: state-of-the-art. In First international meeting on microbial phosphate solubilization. *Springer, Dordrecht* (pp. 229-238). [https://doi.org/10.1007/978-1-4020-5765-6\\_34](https://doi.org/10.1007/978-1-4020-5765-6_34)
- Jannathulla, R., Dayal, J.S., Vasanthakumar, D., Ambasankar, K. and Muralidhar, M. (2017). Effect of fermentation methods on amino acids, fiber fractions and anti-nutritional factors in different plant protein sources and essential amino acid index for *Penaeus vannamei* Boone 1931. <https://doi.org/10.21077/ijf.2017.64.2.60341-07>
- Jia, X.H., Zhang, C.L., Shi, D.J., Zhuang, M.M., Wang, X., Jia, R., Zhang, Z.Y., Huang, J., Sun, Y.H., Qian, W.Y. and Peng, G.H. (2016) Oral administration of Anabaena-expressed VP28 for both drug and food against white spot syndrome virus in shrimp. *Journal Appl Phycol* 28(2):1001-9.
- Kaiser, D., Unger, D., Qiu, G., Zhou, H. and Gan, H. (2013). Natural and human influences on nutrient transport through a small subtropical Chinese estuary. *The Science of the Total Environment* 450-451, 92–107. <https://doi.org/10.1016/j.scitotenv.2013.01.096>
- Kanther, M., Sun, X., Mhlbauer, M., Mackey, L.C., Flynn, E.J. and Bagnat, M. (2011). Microbial colonization induces dynamic temporal and spatial patterns of NF-B activation in the zebrafish digestive tract. *Gastroenterol* 141:197-207. <https://doi.org/10.1053/j.gastro.2011.03.042>
- Karthiga Rani, M., Chelladurai, G. and Jayanthi, G.(2016). Isolation and identification of bacteria from marine market fish *Scomberomorus guttatus* (Bloch and Schneider, 1801) from Madurai district, Tamil Nadu, India. *Journal Parasit Disease* 40(3):1062-1065. <https://doi.org/10.1007/s12639-014-0634-0>
- Kesarcodi-Watson, A., Kaspar, H., Lategan, M.J. and Gibson, L. (2008) Probiotics in aquaculture: the need, principles and mechanisms of action and screening processes. *Aquac* 274(1):1-14. <https://doi.org/10.1016/j.aquaculture.2007.11.019>
- Kim, D.H. and Austin, B. (2006). Innate immune responses in rainbow trout (*Oncorhynchus mykiss*, Walbaum) induced by probiotics. *Fish Shellfish Immunology* 21(5):513-524. <https://doi.org/10.1016/j.fsi.2006.02.007>
- Kim, J.H., Kim, S.K. and Kim, J.H. (2018). Bio-floc technology application in flatfish *Paralichthys olivaceus* culture: Effects on water quality, growth, hematological parameters, and immune responses. *Aquac* 495:703-709. <https://doi.org/10.1016/j.aquaculture.2018.06.034>
- LeBlanc, J.G., Chain, F., Martin, R., Bermudez-Humaran, L.G., Courau, S. and Langella, P. (2017). Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by commensal and probiotic bacteria. *Microb Cell Fact* 16:79. <https://doi.org/10.1186/s1234-017-0691-z>
- LeBlanc, J.G., Milani, C., De Giori, G.S., Sesma, F., Van Sinderen, D. and Ventura, M. (2013). Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnology* 24(2):160-168. <https://doi.org/10.1016/j.copbio.2012.08.005>
- Li, T., Li, H., Gatesoupe, F.J., She, R., Lin, Q., Yan, X., Li, X. et al. (2017). Bacterial signatures of "Red-Operculum" disease in the gut of crucian carp (*Carassius auratus*). *Microb Ecol* 74(3):510-521 <https://doi.org/10.1007/s00248-017-0967-1>
- Li, T., Long, M., Ji, C., Shen, Z., Gatesoupe, F. J., Zhang, X., & Li, A. (2016). Alterations of the gut microbiome of largemouth bronze gudgeon (*Coreius guichenoti*) suffering from furunculosis. *Scientific reports*, 6(1), 30606.
- Limbu, S.M., Zhou, L., Sun, S.X., Zhang, M.L. and Du, Z.Y. (2018). Chronic exposure to low environmental concentrations and legal aquaculture doses of antibiotics cause systemic adverse effects in Nile tilapia and provoke differential human health risk. *Environment International* 115:205-219. <https://doi.org/10.1016/j.envint.2018.03.034>. Epub 2018 Mar 28
- Martinez C.P., Ibáñez, A.L., Monroy Hermosillo, O.A. and Ramirez Saad, H.C. (2012). Use of probiotics in aquaculture. *International Sch Research Notices* 5(1):55-59. <https://doi.org/10.5402/2012/916845>
- Merrifield, D.L., Bradley, G., Baker, R.T.M. and Davies, S.J. (2010). Probiotic applications for rainbow trout (*Oncorhynchus mykiss*, Walbaum) II. Effects on growth performance, feed utilization, intestinal microbiota and related health criteria postantibiotic treatment. *Aquac Nutr* 16(5):496-503. <https://doi.org/10.1111/j.1365-2095.2009.00688.x>

- Merrifield, D.L., Dimitroglou, A., Bradley, G., Baker, R.T.M. and Davies, S.J. (2010) Probiotic applications for rainbow trout (*Oncorhynchus mykiss*, Walbaum) I. Effects on growth performance, feed utilization, intestinal microbiota and related health criteria. *Aquac Nutr* 16(5), 504-510. <https://doi.org/10.1111/j.1365-2095.2009.00689.x>
- Mocanu, E.E., Savin, V., Popa, M.D. and Dima, F.M. (2022). The Effect of Probiotics on Growth Performance, Haematological and Biochemical Profiles in Siberian Sturgeon (*Acipenser baerii* Brandt, 1869). *Fishes* 7(5):239. <https://doi.org/10.3390/fishes7050239>
- Mujeeb R.K.M., Jesmi, Y., Thomas, A.P. and Mohamed Hatha, A.A. (2010). Probiotic effect of *Bacillus* NL110 and *Vibrio* NE17 on the survival, growth performance and immune response of *Macrobrachium rosenbergii* (de Man). *Aquac Research* 41(9): 120-134. <https://doi.org/10.1111/j.1365-2109.2009.02473.x>
- Nakano, T. (2007). Microorganism. En dietary supplements for the health and quality of cultured fish. *CAB International*, London, UK.
- Nie, L., Zhou, Q.J., Qiao, Y. and Chen, J. (2017). Interplay between the gut microbiota and immune responses of ayu (*Plecoglossus altivelis*) during *Vibrio anguillarum* infection. *Fish Shellfish Immunol* 68:479-487. <https://doi.org/10.1016/j.fsi.2017.07.054>
- Nikoskelainen, S., Ouweland, A.C., Bylund, G., Salminen, S. and Lilius, M. (2003) Immune enhancement in rainbow trout (*Oncorhynchus mykiss*) by potential probiotic bacteria (*Lactobacillus rhamnosus*). *Fish Shellfish Immunol* 15(5):443-452. [https://doi.org/10.1016/S1050-4648\(03\)00023-8](https://doi.org/10.1016/S1050-4648(03)00023-8)
- Nimrat, S., Boonthai, T. and Vuthiphandchai, V. (2011). Effects of probiotic forms, compositions of and mode of probiotic administration on rearing of Pacific white shrimp (*Litopenaeus vannamei*) larvae and postlarvae. *Animal Feed Science Technology* 169(3-4):244-258. <https://doi.org/10.1016/j.anifeeds.2011.07.003>
- Omenwa, V.C., Mbakwem-Aniebo, C. and Ibiene, A.A. (2015). Effects of selected probiotics on the growth and survival of fry-fingerlings of *Clarias gariepinus*. *Pharm Biology Science* 10:89-93. <http://dx.doi.org/10.9790/3008-10518993>
- Panigrahi, A., Kiron, V., Kobayashi, T., Puangkaew, J., Satoh, S. and Sugita, H. (2004). Immune responses in rainbow trout *Oncorhynchus mykiss* induced by a potential probiotic bacteria *Lactobacillus rhamnosus* JCM 1136. *Vet Immunol Immunopathol* 102(4):379-388. doi: 10.1016/j.vetimm.2004.08.006.
- Panigrahi, A., Kiron, V., Satoh, S., Hirono, I., Kobayashi, T., Sugita, H., & Aoki, T. (2007). Immune modulation and expression of cytokine genes in rainbow trout *Oncorhynchus mykiss* upon probiotic feeding. *Developmental & Comparative Immunology*, 31(4), 372-382.
- Perera, W.M.K., Carter, C.G. and Houlihan, D.F. (1995). Feed consumption, growth and growth efficiency of rainbow trout (*Oncorhynchus mykiss* Walbaum) fed on diets containing a bacter Arnold ial single-cell protein. *Br J Nutr* 73(4):591-603. <https://doi.org/10.1079/bjn19950061>
- Picchiatti, S., Fausto, A.M., Randelli, E., Carnevali, O., Taddei, A.R., Buonocore, F. and Abelli, L. (2009) Early treatment with *Lactobacillus delbrueckii* strain induces an increase in intestinal T-cells and granulocytes and modulates immune-related genes of larval *Dicentrarchus labrax* (L.). *Fish shellfish immunol* 26(3):368-376 <https://doi.org/10.1016/j.fsi.2008.10.008>
- Pirarat, N., Kobayashi, T., Katagiri, T., Maita, M., & Endo, M. (2006). Protective effects and mechanisms of a probiotic bacterium *Lactobacillus rhamnosus* against experimental *Edwardsiella tarda* infection in tilapia (*Oreochromis niloticus*). *Veterinary immunology and immunopathology*, 113(3-4), 339-347.
- Ravi, A.V., Musthafa, K.S., Jegathambal, G., Kathiresan, K. and Pandian, S.K. (2007). Screening and evaluation of probiotics as a biocontrol agent against pathogenic *Vibriosis* in marine aquaculture. *Lett Appl Microbiol* 45(2):219-223. <https://doi.org/10.1111/j.1472765X.2007.02180.x>
- Reid, G., Sanders, M., Gaskins, H.R., Gibson, G.R., Mercenier, A., Rastall, R., Roberfroid, M., Rowland, I., Cherbut, C. and Klaenhammer, T.R. (2003) New scientific paradigms for probiotics and prebiotics. *Journal Clin Gastroenterol* 37:105-118. <https://doi.org/10.1097/00004836-200308000-00004>
- Robertson, P.A.W., O'Dowd, C., Burrells, C., Williams, P. and Austin, B. (2000). Use of *Carnobacterium* sp. as a probiotic for Atlantic salmon (*Salmo salar* L.) and rainbow trout (*Oncorhynchus mykiss*, Walbaum). *Aquac* 185(3-4):235-243. [https://doi.org/10.1016/S0044-8486\(99\)00349-X](https://doi.org/10.1016/S0044-8486(99)00349-X)
- Rolig, A.S., Sweeney, E.G., Kaye, L.E., DeSantis, M.D., Perkins, A., Banse, A.V. and Guillemain, K. (2018). A bacterial immunomodulatory protein with lipocalin-like domains facilitates host-bacteria mutualism in larval zebrafish. *Elife*, 7:37172. <https://doi.org/10.7554/eLife.37172>
- Romarheim, O.H., Øverland, M., Mydland, L.T., Skrede, A. and Landsverk, T. (2011). Bacteria grown on natural gas prevent soybean meal-induced enteritis in Atlantic salmon. *Journal Nutrition* 141(1):124-130. <https://doi.org/10.3945/jn.110.128900>
- Ronson, P.J. and Medina, R. (2002) Probióticos en la Acuicultura. *Ciencia y Mar* Notas.
- Salinas, I., Cuesta, A., Esteban, M.Á. and Meseguer, J. (2005). Dietary administration of *Lactobacillus delbrueckii* and *Bacillus subtilis*, single or combined, on gilthead seabream cellular innate immune responses. *Fish Shellfish Immunol* 19(1):67-77. <https://doi.org/10.1016/j.fsi.2004.11.007>
- Salinas, I., Díaz-Rosales, P., Cuesta, A., Meseguer, J., Chabrilón, M., Morinón, M.A. and Esteban, M.A. (2006) Effect of heat-inactivated fish and non-fish derived probiotics on the innate immune parameters of a teleost fish (*Sparus aurata* L.). *Veterinary Immunol Immunopathol* 111(3-4):279-286. <https://doi.org/10.1016/j.vetimm.2006.01.020>
- Sharifuzzaman, S. M. and Austin, B. (2009). Influence of probiotic feeding duration on disease resistance and immune parameters in rainbow trout. *Fish & Shellfish Immunology*, 27(3), 440-445. <https://doi.org/10.1016/j.fsi.2009.06.010>
- Sunitha, K. and Krishna, P.V. (2016). Efficacy of probiotics in water quality and bacterial biochemical characterization of fish ponds. *International Journal Curr Microbiology Appl Science* 5(9): 30-37 <http://dx.doi.org/10.20546/ijcm.2016.509.004>
- Tan, H.Y., Chen, S.W. and Hu, S.Y. (2019) Improvements in the growth performance, immunity, disease resistance, and gut microbiota by the probiotic *Rummeliibacillus stabekisii* in Nile tilapia (*Oreochromis niloticus*). *Fish Shellfish Immunol* 92:265-275. <https://doi.org/10.1016/j.fsi.2019.06.027>
- Taoka, Y. and Maeda Jo, J.Y. (2006). Growth, stress tolerance and non-specific immune response of Japanese flounder *Paralichthys olivaceus* to probiotics in a closed recirculating system. *Fish Science* 72(2):310-321. <https://doi.org/10.1111/j.14442906.2006.01152.x>

- Tarnecki, A. M., Wafapoor, M., Phillips, R. N., & Rhody, N. R. (2019). Benefits of a *Bacillus* probiotic to larval fish survival and transport stress resistance. *Scientific reports*, 9(1), 4892.
- Thatcher, C., Høj, L. and Bourne, D. G. (2022). Probiotics for coral aquaculture: challenges and considerations. *Current Opinion in Biotechnology*, 73, 380–386. <https://doi.org/10.1016/j.copbio.2021.09.009>
- Teshima, S., & Kashiwada, K. (1967). Studies on the production of B vitamins by intestinal bacteria of fish. III. Isolation of vitamin B12 synthesizing bacteria and their bacteriological properties. *Bull. Jap. Soc. Sci. Fish*, 33, 979-83.
- Thorpe, A. and Castillo, C.Z. (2018). The economic value of inland fisheries. In *Review of the state of the world fishery resources: inland fisheries* (pp. 214-253). Food and Agriculture Organization of the United Nations. <http://www.fao.org/3/CA0388EN/ca0388en.pdf>
- Tovar, D., Zambonino, J., Cahu, C., Gatesoupe, F.J., Vázquez-Juárez, R. and Lésel, R. (2002). Effect of live yeast incorporation in compound diet on digestive enzyme activity in sea bass (*Dicentrarchus labrax*) larvae. *Aquac* 204(1-2):113-123. [https://doi.org/10.1016/S0044-8486\(01\)00650-0](https://doi.org/10.1016/S0044-8486(01)00650-0)
- Valente, L.M., Araújo, M., Batista, S., Peixoto, M.J., Sousa-Pinto, I., Brotas, V. and Rema, P. (2016) Carotenoid deposition, flesh quality and immunological response of Nile tilapia fed increasing levels of IMTA-cultivated *Ulva* spp. *Journal Appl Phycol* 28(1):691-701. <https://doi.org/10.1007/s10811-015-0590-9>
- Verschuere, L., Rombaut, G., Sorgeloos, P. and Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology Mol Biology Review* 64(4):655-671. <https://doi.org/10.1128/mbr.64.4.655-671.2000>
- Vijayan, K.K., Singh, I.B., Jayaprakash, N.S., Alavandi, S.V., Pai, S.S., Preetha, R. and Santiago, T.C. (2006). A brackishwater isolate of *Pseudomonas* PS-102, a potential antagonistic bacterium against pathogenic vibrios in penaeid and non-penaeid rearing systems. *Aquac* 251(2-4):192-200. <https://doi.org/10.1016/j.aquaculture.2005.10.010>
- Villena, J. and Kitazawa, H. (2020). The Modulation of Mucosal Antiviral Immunity by Immunobiotics: Could They Offer Any Benefit in the SARS-CoV-2 Pandemic?. *Frontiers in Physiology*, 11, 699. <https://doi.org/10.3389/fphys.2020.00699>
- Vine, N.G., Leukes, W.D. and Kaiser, H. (2004). In vitro growth characteristics of five candidate aquaculture probiotics and two fish pathogens grown in fish intestinal mucus. *FEMS Microbiol Lett* 231(1):145-152. [https://doi.org/10.1016/S0378-1097\(03\)00954-6](https://doi.org/10.1016/S0378-1097(03)00954-6).
- Vine, N.G., Leukes, W.D. and Kaiser, H. (2006). Probiotics in marine larviculture. *FEMS Microbiol Review* 30(3):404-427. <https://doi.org/10.1111/j.15746976.2006.00017.x>
- Wang, C., Chuprom, J., Wang, Y. and Fu, L. (2020). Beneficial bacteria for aquaculture: nutrition, bacteriostasis and immunoregulation. *Journal Appl Microbiology* 128(1):28-40. <https://doi.org/10.1111/jam.14383>
- Wang, Y.B., Li, J.R. and Lin, J. (2008). Probiotics in aquaculture: challenges and outlook. *Aquac* 281(1-4):1-4. <http://dx.doi.org/10.1016/j.aquaculture.2008.06.002>
- Wanka, K.M., Damerau, T., Costas, B., Krueger, A., Schulz, C. and Wuertz, S. (2018). Isolation and characterization of native probiotics for fish farming. *Bmc Microbiology* 18(1):1-13. <https://doi.org/10.1186/s12866-018-1260-2>
- World Health Organization (2006). Report of a joint FAO/OIE/WHO Expert Consultation on antimicrobial use in aquaculture and antimicrobial resistance, *Seoul, Republic of Korea*, 13–16. <https://apps.who.int/iris/handle/10665/133869>
- Xu, H.M., Rong, Y.J., Zhao, M.X., Song, B. and Chi, Z.M. (2014). Antibacterial activity of the lipopeptides produced by *Bacillus amyloliquefaciens* M1 against multidrug-resistant *Vibrio* spp. isolated from diseased marine animals. *App Microbiology Biotechnol*, 98(1):127–36. [10.1007/s00253-013-5291-1](https://doi.org/10.1007/s00253-013-5291-1)
- Yanbo, W. and Zirong, X. (2006). Effect of probiotics for common carp (*Cyprinus carpio*) based on growth performance and digestive enzyme activities. *Animal Feed Science Technology* 127(3-4): 283-292. <https://doi.org/10.1016/j.anifeedsci.2005.09.003>
- Yukgehnash, K., Kumar, P., Sivachandran, P., Marimuthu, K., Arshad, A., Paray, B.A. and Arockiaraj, J. (2020). Gut microbiota metagenomics in aquaculture: factors influencing gut microbiome and its physiological role in fish. *Review Aquac* 12:1903–1927. <https://doi.org/10.1111/raq.12416>
- Zhou, X.X., Wang, Y.B. and Li, W.F. (2009). Effect of probiotics on larvae shrimp (*Penaeus vannamei*) based on water quality, survival rate and digestive enzyme activities. *Aquac* 287(3-4):349-353. <https://doi.org/10.1016/j.aquaculture.2008.10.046>