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The Potential of Araneae as Biological Control Agents against Honey-wax Pests (Pyralidae)

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ABSTRACT

A diverse array of spider species inhabits agroecosystems and wild mountainsides, where beekeeping boxes are temporarily installed for natural feeding. Over the past decade, the wax moths *Achroia grisella* and *Galleria mellonella* have become widespread pests in bee colonies within Armenian agrosystems and have also caused damage to stored honeybee combs. Spiders, as predators, employ various strategies to eliminate pests in nature and beekeeping boxes, acting as effective biological control agents. This study aimed to evaluate the functional responses of several Aranea species to these pests, providing viable options for biological control technologies without pesticide use. In this research, eleven spider species were tested, and different functional responses were observed against wax moth larvae and adults. The findings highlighted that *Steatoda paykulliana* was the most significant natural enemy and *Thanatus pictus* was the least effective predator. In addition, intraspecific interference among the eleven spider species demonstrated that increased spider density led to a decreased predation ratio. The results further signified a positive correlation between predator density and the intensity of scrambling competition.

Keywords: Araneae, Biocontrol, Functional response, Pyralidae.

INTRODUCTION

The wax moth species greater *Galleria mellonella* and lesser *Achroia grisella* (Lepidoptera: Pyralidae) are pests of honeybee (*Apis mellifera*) colonies that are distributed worldwide. Their larvae can cause extensive damage to stored wax combs when the bee colony is weak, especially in stressed colonies (Ellis et al. 2013), and transmit incurable and noneliminated viral pathogens (Kwadha et al. 2017). Strong colonies are more preferred by female moths (Williams, 1997). The wax moth is efficient at degrading polyethylene, the most common form of plastic (Bombelli et al. 2017), and can inspire the development of technologies to avoid plastic pollution.

Natural ecosystem and agrosystem pests have been controlled by the indiscriminate use of chemical insecticides for decades. The immense reliance on pesticides has resulted in biodiversity loss, toxicity issues, and widespread environmental pollution (Brühll & Zaller, 2019). The circulation of natural predators to control pest dynamics is one of the various efforts that have been made recently to combine or even substitute insecticides in the management of integrated or invasive pests (Miller et al. 2006; Di et al. 2021).

Spiders (Araneae) are one of the diverse groups of animals in the world and include 50,105 species according to the World Spider Catalog (World Spider Catalog, 2024). Hence, members of this group are now being used as predators in agrosystem to decrease the use of chemical pesticides, thereby contributing to ecological sustainability.

Several advanced studies have used Spider populations to control pests (Fagan & Hurd, 1991; Hayes & Lockley, 1990). Nyffeler et al. were pioneers in studying the ability of Lycosidae spiders to manage cotton field pests of

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A Publication of Unique Scientific Publishers the Noctuidae family and the potential of black widow spiders in tackling wheat field pests (Nyffeler & Benz, 1988; Nyffeler et al. 1992). Young and Edwards studied a few spider species in the USA as predators of crop pests (Young & Edwards, 1990). Furthermore, researchers have revealed the positive effects of spiders in controlling various pests (Marc et al. 1999; Snyder & Wise, 1999) and many investigations have reported encouraging results (Huang et al. 2018; Matteson, 2000). All the previous studies have attempted to represent their outcome around the predator and pest numbers that would be tantamount to the processes in the field or nature (Rodrigues et al. 2013).

This research aimed to evaluate the role of spiders as generalist predators in the biological control of wax moths in beekeeping equipment and to quantify their predation efficiency. In addition, this study intended to identify specific spider species with the highest potential for effective pest management and to understand the behavioral interactions between spiders and wax moths. Furthermore, this investigation sought to evaluate the impact of spider predation on the overall health and productivity of beekeeping operations and to explore the potential for integrating spider-based biocontrol methods with other pest management strategies. This attempt could provide beekeepers with novel approaches for the biological control of pests and aid in healthy product management.

Spider Fauna of Armenia

According to WSC (2024), Armenia is home to 229 spider species belonging to 34 families. Various researchers have documented the spider fauna of Armenia and have contributed to our understanding of its arachnofauna (Ovtsharenko, 1994; Tanasevitch, 1990; Dunin, 1992; Marusik, 1989; Logunov, 2015; Logunov & Guseinov, 2002; Rakov & Logunov, 1997; Dunin & Zacharjan, 1991; Mikhailov, 2000, 2013, 2016; Mikhailov & Propistsova, 2017). More recent studies have been conducted by Zarikian et al. (Zarikian, 2020, 2021, 2022; Zarikian & Kalashian, 2021; Zarikian et al., 2022, 2023).

MATERIALS & METHODS

In this study, 11 species of Araneae that represented 9 families and 9 genera were used (Table 1). These were collected by handpicking sweeping from various regions of Armenia during the experimental period (2021–2022). The spiders were identified based on the identification keys of "Spiders of Europe" (Nentwig et al. 2024). They were installed in plastic boxes ($58 \times 37 \times 16$ cm) (a separate box for each species) in which a single infected wax comb was laid (infected by both moth species arbitrarily). The laboratory conditions were $26 \pm 0.5^{\circ}$ C and $60 \pm 3\%$ relative humidity.

The spiders were starved for 48h, and then, one adult spider of each species (only female spiders were used to avoid cannibalism) was dropped in each box (Fig. 1). Each box contained one spider and larvae (at the third instar stage) of moths at different densities (10, 20, 30, 40, 50,

Table 1: The spiders' species used as biocontrol agents in the experiment.

No.	Family	Species
1	Agelenidae	Tegenaria domestica (Clerck, 1757)
2	Araneidae	**Araneus quadratus Clerck, 1757
3	Dysderidae	**Dysdera crocata C. L. Koch, 1838
4	Lycosidae	Lycosa praegrandisC. L. Koch, 1836
5		**Lycosa singoriensis (Laxmann, 1770)
6	Philodromidae	Thanatus pictus L. Koch, 1881
7	Pisauridae	**Pisaura mirabilis (Clerck, 1757)
8	Salticidae	Philaeus chrysops (Poda, 1761)
9	Scytididae	Scytodes thoracica (Latreille, 1802)
10	Theridiidae	Steatoda paykulliana Walckenaer, 1805
11		**Steatoda dahli (Nosek, 1905)

**First record for Armenian fauna in this paper.



Fig. 1: Steatoda paykulliana installed in an experiment box after starving for 48 h

and 60) (Fig. 2). The experiments were repeated with five replicates per density level. The number of preys that survived after 48 h was recorded. The functional responses of each spider species in each family were analyzed according to the Holling II model (Holling, 1959). The same experiment was performed with adult moths too.

Quantities of larvae and adult moths in each box before and after preying by spiders for 2 days were calculated and recorded in Table 2.

The experiment was continued to determine the intensity of scrambling competition of the spiders toward larvae and adult moths. For this study, spiders from each species were placed in 6 boxes in which 40 larvae (followed by the adults) were laid. There were 1, 2, 3, 4, and 5 spiders per box, with the control box having no spiders (Fig. 3).

The spiders underwent a period of starvation that lasted for 2 days prior to the commencement of the experiment. Each density trial was replicated four times. The count of surviving moths within each enclosure was

Table 2: Functional responses of spiders to larvae and adults of moths (a') the spider's attack coefficient, (N) the density of the moths, (Na) the number of moths (larvae or adults) predated by the spider, (r) the fitting coefficient, (Th,) the spider's handling time per moth, and (N max) the maximum number of moths predated by the spider

Spider species	Life stage of prey	Equation of functional response	r	Th(h)	a'	Nmax
T. domestica	larva	1/Na =3.233/N + 0.072	0.905	0.072	0.309	10
	adult	1/Na =7.99/N + 0.078	0.974	0.078	0.125	6.2
A. quadratus	larva	1/Na =8.951/N + 0.003	0.994	0.003	0.111	13.6
	adult	1/Na =4.139/N + 0.241	0.977	0.031	0.241	17.8
D. crocata	larva	1/Na =4.058/N + 0.017	0.997	0.017	0.246	21.08
	adult	1/Na =22.369/N + 0.102	0.998	0.102	0.044	12.8
L. praegrandis	larva	1/Na =2.407/N + 0.007	0.994	0.007	0.415	40.2
	adult	1/Na =2.726/N + 0.013	0.993	0.013	0.439	32.6
L. singoriensis	larva	1/Na =2.298/N + 0.008	0.988	0.008	0.435	41.8
	adult	1/Na =2.869/N + 0.15	0.977	0.15	0.348	33.2
Th. pictus	larva	1/Na =41.8/N + 0.525	0.928	0.525	0.023	1.8
	adult	1/Na =30.78/N + 0.182	0.970	0.182	0.032	4.2
P. mirabilis	larva	1/Na =4.722/N + 0.024	0.989	0.024	0.211	19
	adult	1/Na =9.393/N - 0.005	0.995	0.005	0.106	16.8
Ph. chrysops	larva	1/Na =3.2/N + 0.033	0.973	0.033	0.312	22.2
	adult	1/Na =2.836/N + 0.33	0.966	0.33	0.352	23.8
S. thoracica	larva	1/Na =53.265/N – 0.212	0.997	0.212	0.018	3.8
	adult	-	0	0	0	0
S. paykulliana	larva	1/Na =2.344/N + 0.008	0.989	0.008	0.426	40.6
	adult	1/Na =1.9016/N + 0.012	0.969	0.012	0.521	42.2
S. dahli	larva	1/Na =3.724/N + 0.028	0.956	0.028	0.268	22
	adult	1/Na =5.171/N + 0.013	0.991	0.013	0.193	20.8



The plastic box where the spiders were installed

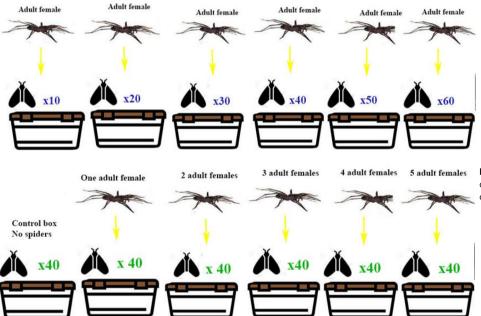


Fig. 2: Graphic explanation of the first part of the experiment

Fig. 3: Graphic explanation of the part of the experiment showing the intensity of spiders' scrambling competition

(1)

recorded after 1 day. Intraspecific competition among spiders for predation on both moth larvae and adults was assessed using the Hassell model (Hassell & Varley, 1969).

The spiders utilized in this study were taxonomically identified and preserved as vouchers in the collections of the Scientific Center of Zoology and Hydroecology at the Institute of the National Academy of Sciences of the Republic of Armenia.

Data Analyses

The Holling type II formula was used to calculate the predatory functional response, as follows:

$$Na = a'TN/(1 + a'ThN)$$

Where, N represents the prey density, Na denotes the number of preys consumed, T indicates the time required for the predator to locate the prey (T = 2 days or 48 h), a' signifies the instantaneous attack rate, and Th represents the time required by the predator to handle one individual, considering a box area of 0.2146 m². The parameters a' and Th were measured using a nonlinear least-squares method. Initially, starting values for a' and Th required for the nonlinear regression (NLR) procedure were determined via linear regression of 1/Na against 1/N. The resulting y-

intercept served as the initial estimate for Th, whereas the reciprocal regression coefficient provided an estimate for a'. Na and N were determined experimentally; on the contrary, a' and Th were derived computationally. Equation (2)

All experimental data regarding the intraspecific disturbance of spider predation on moth larvae and adults were analyzed using the Hassell model, as expressed below:

$$E = QP^{-m} \tag{2}$$

Where, E represents the predation ratio of the spiders, P denotes the spider density, Q signifies the searching constant, and m represents the interference constant. Q and m were estimated using the following equation:

$$E = \frac{N_a}{N \times P} \tag{3}$$

Where, Na represents the aggregate number of moths consumed by all spiders, N denotes the density of moths, and P indicates the density of spiders.

The predation competition among spiders for the prey was characterized using scrambling competition. Hence, the intensity of scrambling competition (I) among spiders was quantified using the following equation:

$$I = \frac{E_1 - E_P}{E_1} \tag{4}$$

Where, E1 represents the predation ratio of a single spider and Ep signifies the predation ratio with a density of spiders denoted by P. The correlation between the intensity of scrambling competition (I) and spider density (P) was derived using the following equation:

$$I = a \lg P + b \tag{5}$$

According to the values of I, the parameters a and b were estimated. All collected data were analyzed using MS Excel 2007 and IBM SPSS statistic 20.

RESULTS

The functional response of the spiders used in this research toward moth larvae and adults was determined. The functional response equations of each species are presented in Table 2 for the 11 species studied. The results indicated that spiders from the same species exhibited varying functional responses to the pest larvae and adults. The fitting coefficients of the 22 equations ranged from 0.905 to 0.998, which implied that these equations were a good fit for all the examined species, with the exception of Scytodes thoracica. Steatoda paykulliana demonstrated the highest ability toward the adults of moths, followed by Lycosa singoriensis toward the larvae. Both S. paykulliana and Lycosa praegrandis exhibited a high reaction toward the larvae, and the highest attack coefficient (0.439) was reached by L. praegrandis toward the pest's adults and the shortest Th (that is, 0.003 h) by Araneus quadratus toward the pest's larvae.

This study examined the functional response of spiders to moth larvae and adults. Table 2 presents the functional response equations for each of the 11 species explored. The findings revealed that spiders belonging to the same species exhibited varying functional responses to pest larvae and adults. The fitting coefficients of the 22 equations ranged from 0.905 to 0.998, which suggested a good fit for all species, except *S. thoracica*. Notably, *S. paykulliana* displayed the highest predation ability against moth adults, followed by *L. singoriensis*, which targeted the larvae. In addition, both *S. paykulliana* and *L. praegrandis* showed strong reactions to larvae. Of these, *L. praegrandis* presented the highest attack coefficient (0.439) against pest adults, and the shortest handling time (0.003 h) was observed for *A. quadratus* when preying upon pest larvae. Thus, *S. paykulliana* was the most important natural enemy, while *Thanathus pictus* had insufficient predatory abilities toward wax moths.

Intraspecific Disturbance and Intensity of Scrambling Competition on Moth Larvae and Adults

The predation ratio was calculated based on Equation (3) and Na values (Table 3), as shown in Table 3. Na and Ep of each spider decreased gradually with the increase in spider density. The searching constant (Q) and the interference constant (m) were calculated with the Hassell model to fit the predator density (Equation 2) using the NLR model (Table 4). In most cases, the fitting coefficients of these equations ranged from 0.797 to 0.990. However, in certain species, this coefficient exceeded 0.990, which alludes that the predation rate of the species was considerably correlated with its density and that the predator density interfered with its predation on moths. This finding denotes that an increase in predator density led to an increase in the interference effect between spider individuals, and as a result, the general predation rate decreased. Significant variations were noted in the Q and m values for both moth larvae and adults. The intensity of scrambling competition, as presented in Table 3, was calculated using Equation (4), whereas the parameters a and b of Equation (5) in Table 5 were estimated using the NRL model. The intensity of scrambling competition was positively correlated with increasing spider density.

DISCUSSION

The wax moths A. grisella and G. mellonella are the major pests of honey wax, and they attack the honeyproducing boxes in most moist areas, especially in territories adjoining forest areas in Armenia. These moths have developed high levels of resistance against pesticides to the extent that using chemical methods is not effective anymore (Cotter et al. 2000). However, the role of natural predators of A. grisella and G. mellonella as spiders in agricultural ecosystems has not been studied extensively. Spiders are the most prevalent predators in the vicinity of beekeeping equipment, and certain species within this group play a significant role in curbing pest populations. Understanding the dynamics between predators (spiders) and pests is a primary objective in global pest management efforts. To implement environmentally safe strategies, various eco-sustainable control methods and integrated pest management programs have recently been evaluated (Zappala et al. 2012).

Table 3: Predation ratio and intensity of scrambling competition of spiders toward two species of moths and their larvae, (Ep) the predation ratio with P density of spiders, (I) the intensity of the scrambling competition, (Na) the total number of pests preved on by all of the spiders in the container

Spider	The life stage of prey	Spider density	Na	Ep	0.000
T. domestica	larva	1 2	5	0.125 0.056	0.000
		3	4.5 4	0.030	0.552 0.736
		4	4.5	0.028	0.776
		5	4.6	0.023	0.816
	adult	1	4	0.1	0.000
		2	3.5	0.043	0.57
		3	3.6	0.03	0.7
		4	4	0.025	0.75
		5	3.8	0.019	0.81
A. quadratus	larva	1	9	0.225	0.000
		2	7.5	0.093	0.586
		3	7.3	0.06	0.733
		4		0.045	0.8
	adult	5 1	6.6 10	0.033 0.250	0.853 0.000
	auuit	2	8	0.230	
		3	8	0.066	0.736
		4	7.5	0.046	0.816
		5	6.8	0.034	0.864
D. crocata	larva	1	13	0.325	0.000
		2	9	0.112	0.655
		3	7.6	0.063	0.806
		4	7.7	0.048	0.852
		5	7.2	0.036	0.889
	adult	1	7	0.175	0.000
		2	5	0.062	0.645
		3	5	0.041	0.765
		4	5.2	0.032	0.817
	lan a	5	4.8	0.024	
L.praegrandis	larva	1	14 0.5	0.350	
		2 3	9.5 o	0.118	0.662
		4	8 8	0.066 0.050	0.811 0.857
		5	7.6	0.038	
	adult	1	11	0.275	0.000
	addit	2	8.5	0.106	0.614
		3	7.3	0.060	0.781
		4	7	0.043	0.843
		5	6.6	0.033	0.880
L. singoriensis	larva	1	17	0.425	0.000
		2		0.131	0.691
		3	9	0.075	0.823
		4		0.048	0.887
	1.11	5	8	0.004	0.990
	adult	1	14	0.350	0.000
		2	9.5		0.662
		3 4	8.3 7.5	0.069 0.046	
		5	7.2	0.040	
Th. Pictus	larva	1	2	0.050	
		2	2	0.025	0.500
		3	2.3	0.019	
		4	2.5	0.015	0.700
		5	2.8	0.014	
	adult	1	3	0.075	0.000
		2	3	0.037	0.506
		3	4	0.033	0.560
		4	4.25		
,		5	4.4	0.022	
P. mirabilis	larva	1	8	0.200	
		2	5.5 6.6	0.068	
		3	6.6	0.055	
		4 5	6.5	0.040	
	adult	5 1	5.8 6	0.029	
	adult	2	6 4.5	0.150 0.056	
		2 3	4.5 6.6	0.056	
		4	5.75		
		5	5.4	0.027	0.820
		-			
Ph.Chrysops	larva	1	12	0.300	0.000

adu	3		6.6	0.055	0.816
adu					
adu			7.5	0.046	0.846
adu	5		6.6	0.033	0.890
			13	0.325	0.000
	2		10.5	0.131	0.596
	3		9	0.075	0.769
	4	ļ	7.75	0.048	0.852
	5	i	6.6	0.033	0.898
S. thoracica larv			1	0.025	0.000
	2		1.5	0.018	0.280
	3		1.3	0.010	0.600
	4	ļ	1.75	0.010	0.600
	5	i	2	0.010	0.600
adu			0	00	0.000
	2		0	00	0.000
	3		0	00	0.000
	4	Ļ	0	00	0.000
	5	i	0	00	0.000
S. paykulliana larv	ra 1		22	0.550	0.000
	2		13.5	0.168	0.694
	3		11.3	0.094	0.829
	4	Ļ	10	0.062	0.887
	5	i	8	0.040	0.927
adu	ılt 1		27	0.675	0.000
	2		15	0.187	0.722
	3		13	0.108	0.840
	4	Ļ	10	0.062	0.908
	5	i	8	0.040	0.940
S. dahli larv	ra 1		11	0.275	0.000
	2		8.5	0.106	0.614
	3		7.6	0.063	0.770
	4	Ļ	6.75	0.042	0.847
	5	i	6	0.030	0.890
adu	ılt 1		9	0.225	0.000
	2		7.5	0.093	0.586
	3		7	0.058	0.742
	4	Ļ	6.25	0.039	0.826
	5		5.6	0.028	0.875

Table 4: The intraspecific disturbance	equations of the spiders: searching				
constant O and interference constant m.					

Spider	Life stage of	Searching	Interference	Fitting
species	prey	constant Q	Constant m	coefficient
T. domestica	larva	0.120	1.065	0.989
	adult	0.095	1.005	0.989
A. quadratus	larva	0.220	1.173	0.998
	adult	0.245	1.221	0.998
D. crocata	larva	0.307	1.365	0.994
	adult	0.162	1.210	0.989
L. praegrandis	larva	0.329	1.380	0.993
	adult	0.269	1.325	0.999
L. singoriensis	larva	0.613	2.421	0.797
	adult	0.335	1.420	0.997
Th. pictus	larva	0.047	0.804	0.984
	adult	0.071	0.732	0.972
P. mirabilis	larva	0.183	1.143	0.975
	adult	0.140	1.005	0.952
Ph. chrysops	larva	0.295	1.384	0.990
	adult	0.337	1.410	0.998
S. thoracica	larva	0.025	0.639	0.901
	adult	0.000	0.000	0.000
S. paykulliana	larva	0.538	1.597	0.998
	adult	0.665	1.724	0.997
S. dahli	larva	0.276	1.366	1.000
	adult	0.227	1.280	0.999

Numerous studies employing the Holling II model have attempted to elucidate the functional response of spiders, yielding results similar to those observed in the current investigation (Maloney et al. 2003; Rao and Kanaujia, 2023). These studies have indicated that as the moth density increases, the spider predation initially rises before reaching a plateau (Lin et al., 2006; Huang et al., 2018). Functional response curves for these spiders typically exhibit the characteristics of type II, wherein prey

Table 5: The equation of intensity of the scrambling-competition for spider's larvae and adults.

species prey equation T. domestica larva I = 0.509 lg P + 0.088 0.92 adult I = 0.494 lg P + 0.093 0.91 A. quadratus larva I = 0.525 lg P + 0.092 0.92	13
adult I = 0.494 lg P + 0.093 0.91	13
···· 5	
A <i>quadratus</i> lanza $I = 0.525 \ln P + 0.092$ 0.92	22
	- 0
adult I = 0.531 lg P + 0.094 0.92	22
D. crocata larva I = 0.548 lg P + 0.116 0.89	92
adult I = 0.526 lg P + 0.114 0.88	38
L. larva I = 0.550 lg P + 0.118 0.88	38
<i>praegrandis</i> adult I = 0.547 lg P + 0.100 0.91	15
<i>L. singoriensis</i> larva I = 0.594 lg P + 0.110 0.91	16
adult I = 0.554 lg P + 0.115 0.89) 5
<i>Th. pictus</i> larva I = 0.449 lg P + 0.078 0.92	23
adult I = 0.424 lg P + 0.079 0.91	14
P. mirabilis larva I = 0.513 lg P + 0.117 0.87	78
adult I = 0.487 lg P + 0.103 0.88	35
Ph. chrysops larva I = 0.556 lg P + 0.099 0.91	14
adult I = 0.558 lg P + 0.089 0.93	35
S. thoracica larva I = 0.408 lg P + 0.025 0.91	17
adult	
S. paykulliana larva I = 0.568 lg P + 0.123 0.88	38
adult I = 0.576 lg P + 0.131 0.87	78
<i>S. dahli</i> larva I = 0.551 lg P + 0.097 0.99	92
adult I = 0.541 lg P + 0.088 0.93	34

consumption peaks with escalating pest density (Mansour & Heimbach, 1993). A plausible explanation lies in the saturation of predation demands once food requirements are met, thus curtailing further predation activity.

Predation escalates with an increase in prey numbers but diminishes with heightened prey density, and a positive correlation exists between I and P (Wu & Wang, 1987). Moreover, predators exhibit reduced predation rates in high-density environments when compared with individual spiders (Pérez-Guerrero et al., 2013).

Intraspecific interference among the 11 spider species under study was assessed using the Hassell model. As the spider density increased, predation ratios declined. Nonetheless, a positive correlation emerged between the intensity of scrambling competition and predator density, which signified a surge in scrambling competition intensity with increasing spider density (Chen et al. 2010).

Conclusion

This research was focused on 11 abundant spiders and investigated their potential to control wax moths. The findings revealed that certain spider species are effective biocontrol agents in wax moth pest management. S. paykulliana, L. singoriensis, and L. praegrandis were highly fierce and exhibited a high predatory rate toward moths; hence, these can be used as pest control agents after evaluating the pest density in the honey-keeping boxes. On the contrary, T. pictus and S. thoracica exhibited the least predatory effects. Intraspecific interactions among the studied spider species were evident. When five spiders were confined to a single enclosure, no aggressive behavior was observed. Nonetheless, an increase in interference among individual spiders led to prolonged prey-searching times and reduced the predation efficiency for each spider.

To validate the applicability of the laboratory findings, the experiment was replicated under natural field conditions, accounting for variables such as activity space, niche overlap, developmental stages, bee community structure, and predator-prey interactions, all of which can impact predator behavior. Laboratory-reared spider populations were introduced into beekeeping environments, where certain colonies were afflicted by the same pest. The field observations closely paralleled those obtained under controlled laboratory settings, albeit with some noticeable variations. These findings will be elaborated upon in a forthcoming publication.

Data Availability Statement

The data presented in this study are available on request from the corresponding author. The voucher material was deposited in the invertebrate collection of NAS RA Scientific Centre of Zoology and Hydro-ecology Institute.

Declaration of Competing Interest

The authors declare that they have no conflict of interest.

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Author's Contribution

Conceptualization, R.P.; methodology, L.H.; formal analysis, M.V.; investigation M.R.; writing & reviewing N.Z; editing, K.A.; visualization, R.P., and R.H.; data collection, M.M., and Z.D. All authors have read and agreed to the published version of the manuscript.

REFERENCES

- Bombelli, P., Howe, C. J., and Bertocchini, F. (2017). Polyethylene biodegradation by larva of the wax moth Galleria mellonella. *Current Biology*, 27, 283–293. <u>https://doi.org/10.1016/j.cub.2017.02.060</u>
- Brühll, C.A., and Zaller, J.G. (2019). Biodiversity Decline as a Consequence of an Inappropriate Environmental Risk Assessment of Pesticides. Frontiers in Environmental Science, 7, 177. https://doi.org/10.3389/fenvs.2019.001 77
- Chen, L., Chen, F., and Chen, L. (2010). Qualitative analysis of a predatorprey model with Holling type Ilfunctional response incorporating a constant prey refuge. *Nonlinear Analysis: Real World Applications*, 11(1), 246–252.
- Cotter, G., Doyle, S., and Kavanagh, K. (2000). Development of an insect model for the in vivo pathogenicity testing of yeasts. *FEMS Immunology and Medical Microbiology*, 27, 163–169.
- Di, N., Zhang, K., Xu, Q., Zhang, F., Harwood, J.D., Wang, S., and Desneux, N. (2021). Predatory ability of *Harmonia axyridis* (Coleoptera: Coccinellidae) and *Orius sauteri* (Hemiptera: Anthocoridae) for suppression of fall armyworm Spodoptera frugiperda (Lepidoptera: Noctuidae). *Insects*, 12, 1063. <u>https://doi.org/10.3390/insects12121063</u>
- Dunin, P.M. (1992). The spider family Dysderidae of the Caucasian fauna (Arachnida Aranei Haplogynae). Arthropoda Selecta, 1 (3), 35-76.
- Dunin, P.M., and Zacharjan, V.A. (1991). New spider species of genus Zodarion from the Caucasus (Aranei, Zodariidae). *Zoological Journal*, 70, 142-144
- Ellis, E.C., Kaplan, J.O., Fuller, D.Q., Vavrus, S., Goldewijk, K.K., and Verburg, P.H. (2013). Used planet: A global history. *Proceedings of the National Academy of Sciences of the United States of America*, 110(20), 7978-7985. <u>http://dx.doi.org/10.1073/pnas.1217241110</u>
- Fagan, W.F., and Hurd, L.E. (1991). Direct and indirect effects of generalist predators on a terrestrial arthropod community. *American Midland Naturalist*, 126, 380–384. <u>https://doi.org/10.1016/0040-809(89)</u> <u>90037-3</u>

- Hassell, M.P., and Varley, G.C. (1969). New inductive population model for insect parasites and its bearing on biological control. *Nature*, 223, 1133–1137.
- Hayes, J.L., and Lockley, T.C. (1990). Prey and nocturnal activity of wolf spiders (Araneae: Lycosidae) in cotton fields in the delta region of Mississippi. *Environmental Entomology*, 19, 1512–1518.
- Holling, C.S. (1959). Some characteristics of simple type of predation and parasitism. *Canadian Entomology*, 91, 385–398.
- Huang, X., Quan, X., Wang, X., Yun, Y., and Peng, Y. (2018). Is the spider a good biological control agent for *Plutella xylostella* (Lepidoptera: Plutellidae)? *Zoologia*, 35, e23481. <u>https://doi.org/10.3897/zoologia.35.</u>
- Kwadha, C. A., Ong'amo, G.O., Ndegwa, P.N., Raina, S.K., and Fombong, A.T. (2017). The Biology and Control of the Greater Wax Moth, *Galleria* mellonella. Insects, 8(2), 61, <u>https://doi.org/10.3390/insects8020061</u>
- Logunov, D.V. (2015). Taxonomic-faunistic notes on the jumping spiders of the Mediterranean (Aranei: Salticidae). *Arthropoda Selecta*, 24, 33-85. <u>https://doi.org/10.15298/arthsel.24.1.03</u>
- Logunov, D.V., and Guseinov, E.F. (2002). Faunistic review of the jumping spiders of Azerbaijan (Aranei: Salticidae), with additional faunistic records from neighboring Caucasian countries. *Arthropoda Selecta*, 10, 243-260.
- Lin, W., Zhang, J., and Lu, Y. (2006). Predation of female adults of *Erigonidium graminicolum* on larvae of *Plutella xylostella* and *Pieris rapae*. Acta Agriculturae Zhejiangensis, 18, 216–220.
- Maloney, D., Drummond, F.A., and Alford, R. (2003). Spider Predation in Agroecosystems: Can Spiders Effectively Control Pest Populations? Technical Bulletin 190, Maine Agricultural and Forest Experiment Station, The University of Maine, pp: 1-32.
- Mansour, F., and Heimbach, U. (1993). Evaluation of lycosid, micryphantid, and linyphiid spiders as predators of *Rhopalosiphum padi* (Hom.: Aphididae) and their functional response to prey density-laboratory experiments. *Entomophaga*, 38(1), 79–87. https://doi.org/10.1007/BF02373142
- Marc, P., Canard, A., and Ysnel, F. (1999). Spiders (Araneae) useful for pest limitation and bioindication. *Agriculture, Ecosystems & Environment*, 74, 229-273.
- Marusik, Y.M. (1989). New data on the fauna and synonyms of the spiders of the USSR (Arachnida, Aranei). In: Lange AB (ed.) Fauna i Ekologiy Paukov i Skorpionov, Arakhnologicheskii. Sbornik Akademia Nauk SSSR, Moscow. 39-52.
- Matteson, P.C. (2000). Insect Pest Management in Tropical Asian Irrigated Rice. Annual Review of Entomology, 45, 549-574. https://doi.org/10.1146/annurev.ento.45.1.549
- Mikhailov, K.G. (2000). Catalogue of the spiders of the of the territories of the former Soviet Union (Arachnida, Aranei). Addendum 3. Zoological Museum of the Moscow State University. 33 pp.
- Mikhailov, K.G. (2013). The spiders (Arachnida: Aranei) of Russia and adjacent countries: a non-annotated checklist. Arthropoda Selecta, Supplement, 3, 1-262.
- Mikhailov, K.G. (2016). Advances in the study of the spider fauna (Aranei) of Russia and adjacent regions: a 2015 update. *Vestnik Zoologii*, 50, 309-320. <u>https://doi.org/10.1515/vzoo-2016-0038</u>.
- Mikhailov, K.G., and Propistsova, E.A. (2017). On the spiders (Arachnida: Aranei) from Armenia. Arthropoda Selecta, 26, 369-371. https://doi.org/10.15298/arthsel.26.4.11
- Miller, D.A., Grand, J.B., and Fondell, T.F. (2006). Anthony, M. Predator functional response and prey survival: Direct and indirect interactions affecting a marked prey population. *Journal of Animal Ecology*, 75, 101–110.
- Nentwig, W., Blick, T., Bosmans, R., Gloor, D., Hänggi, A., and Kropf, C. (2024). Spiders of Europe. Version 2024. <u>https://doi.org/10.24436/1</u>
- Nyffeler, M., and Benz, G. (1988). Prey analysis of the spider Achaearanea riparia (Black w.) (Araneae, Theridiidae), a generalist predator in winter wheat fields. *Journal of Applied Entomology*, 106, 425-431.

- Nyffeler, M., Sterling, W.L., and Dean, D.A. (1992). Impact of the striped lynx spider (Araneae: Oxyopidae) and other natural enemies on the cotton feahopper (Hemiptera: Miridae) in Texas cotton. *Environmental Entomology*, 2I, 1178-1188.
- Ovtsharenko, V.I. (1994). The remarkable montane distribution of ground spiders of the genus Parasyrisca (Gnaphosidae) in the Holarctic. *American Arachnology*, 50, 7-8.
- Pérez-Guerrero, S., Gelan-Begna, A., Tamajón, A., and Vargas-Osuna, A. (2013). Potential predation of non-webbuilding spider assemblage on cotton pests *Helicoverpa armigera and Spodoptera littoralis* (Lepidoptera: Noctuidae). *Biocontrol Science and Technology*, 23(3), 335–347. https://doi.org/10.1080/09583157.2012.758237
- Rakov, S.Y., and Logunov, D.V. (1997). A critical review of the genus Heliophanus C. L. Koch, 1833, of Middle Asia and the Caucasus (Aranei Salticidae). Arthropoda Selecta, 5(3/4), 67-104.
- Rao, K., and Kanaujia, A. (2023). Spiders' Impact on Controlling Insect Pests in the Agricultural Ecosystem. *International Journal of Research Publication and Reviews*, 14(7).
- Rodrigues, E.N.L., Mendonça, Jr. M. S., Fritz, L.L., Heinrichs, E.A., and Fiuza, L. (2013). Effect of the insecticide Lambda-cyhalothrin on rice spider populations in Southern Brazil. *Zoologia* (Curitiba), 30(6), 615-622. <u>https://doi.org/10.1590/S1984-4670201300</u>
- Snyder, W.E., and Wise, D.H. (1999). Predator interference and the establishment of generalist predator populations for biocontrol. *Biological Control*, 15, 283-292.
- Tanasevitch, A.V. (1990). The spider family Linyphiidae in the fauna of the Caucasus (Arachnida, Aranei). In: B. R. Striganova (ed.) Fauna nazemnykh bespozvonochnykh Kavkaza. Moscow, Akaedemia Nauk, pp. 5-114.
- Williams, J.L. (1997). Insects: Lepidoptera (moths). In *Honey Bee Pests, Predators, and Diseases*; Morse, R., Flottum, K., Eds.; Al Root Company: Medina, OH, USA, pp. 121–141.
- World Spider Catalog (2024). World spider catalog. Version 21.5. Natural History Museum, Bern. <u>https://wsc.nmbe.ch</u>
- Wu, L.L., and Wang, H.Q. (1987). Functional response for a predator *Pirata* subpiraticus (Araneae: Lycosidae) to its prey. *Chinese Journal of Biological Control*, 3, 7–10.
- Young, O.P., and Edwards, G.B. (1990). Spiders in United States field crops and their potential impact on crop pests. *Journal of Arachnology*, 18, 1–27.
- Zappala, L., Bernardo, U., Biondi, A., Cocco, A., Deliperi, S., Delrio, G., Giorgini, M., Pedata, P., Repisarda, C., Garzia, G.T., and Siscaro, G. (2012). Recruitment of native parasitoides by the exotic pest, Tuta abusoluta in Sourthern Eatly. *Bull Insectol*, 65(01), 51-61.
- Zarikian, N. (2020). A Contribution to the Checklist of the Jumping Spiders (Araneae: Salticidae) of Armenia. *Bulletin of the Iraq Natural History Museum*, 16(2), 193-202. <u>https://doi.org/10.26842/binhm.7.2020.16.2</u>, 0193
- Zarikian, N.A. (2021). A survey of running crab spiders Philodromidae (Araneae) of Armenia. *Bulletin of the Iraq Natural History Museum*, 16(4), 495-508. <u>https://doi.org/10.26842/binhm.7.2021.16.4.0495</u>
- Zarikian, N. A. (2022). New records of Araneae from Armenia. Indian Journal of Entomology, 84(4), 757-760. <u>https://doi.org/10.55446/IJE.2022.688</u>
- Zarikian, N., and Kalashian, M. (2021). An annotated checklist of spiders deposited in the Arachnida collection of the Institute of Zoology, Scientific Center of Zoology and Hydroecology of the NAS RA, Yerevan, Armenia. Part I, Arachnologische Mitteilungen, 61(1), 11-19. https://doi.org/10.30963/aramit6102
- Zarikian, N.A., Propistsova, E.A., and Marusik, Y.M. (2022). On spider families (Arachnida: Araneae) new to Armenia. *Israel Journal of Entomology*, 51(2021), 103-117. <u>https://doi.org/10.5281/zenodo.6466083</u>
- Zarikian, N., Dilbaryan, K., Khachatryan, A. and Harutyunova, L. (2023). Species composition and diversity of spider (Arachnida: Araneae) in the northern forests of Armenia, *Biodiversity*, 24(1-2), 66-75. <u>https://doi.org/10.1080/14888386.2023.2184424</u>